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(54) Title: CHIMERIC MOLECULES CONTAINING A MODULE ABLE TO TARGET SPECIFIC CELLS AND A MODULE REGULATING THE APOPTOGENIC FUNCTION OF THE PERMEABILITY TRANSITION PORE COMPLEX (PTPC)

(57) Abstract: A chimeric polypeptide has the formula: pTox-pTarg, wherein pTox is a viral apoptotic peptide, such as the Vpr peptide of HIV-1 or a fragment of the Vpr peptide of HIV-1 containing the amino acid motif H(F/S)RIG that interacts with mitochondrial inner membrane, adenine nucleotide translocation (ANT) protein of a cell. pTarg is an antibody or an antibody fragment that binds to the outer membrane of the cell. Binding of the chimeric polypeptide to the cell is followed by apoptosis of the cell. A vector encoding a chimeric polypeptide and a recombinant host cell comprising the vector are provided. The chimeric polypeptide is useful for targeting pTox to cells, such as cancer cells.



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CHIMERIC MOLECULES CONTAINING A MODULE ABLE TO TARGET
SPECIFIC CELLS AND A MODULE REGULATING THE APOPTOGENIC
FUNCTION OF THE PERMEABILITY TRANSITION PORE COMPLEX (PTPC)

CROSS-REFERENCE TO RELATED APPLICATIONS

The application hereby claims the benefit under 35 U.S.C. § 119(e) of United States provisional application Serial No. 60/265,594, filed February 2, 2001. The entire disclosure of this application is relied upon and incorporated by reference herein.

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to cell death regulatory molecules for therapeutic use. More specifically, this invention relates to molecules in which a peptidic or pseudo-peptidic part acting on the permeability transition pore complex (PTPC) is covalently linked to cell-targeting molecules including antibodies, recombinant antibody fragments or homing peptides. The resulting chimeric molecules are polypeptides or peptidomimetic molecules which target the PTPC and/or its major component the adenine nucleotide translocation (ANT) to induce or inhibit cell death (apoptosis). This invention also relates to such chimeric molecules when the PTPC-interacting part is an apoptogenic HIV-1 Vpr-derived peptide (or pseudopeptide) or an ANT-derived peptide (or pseudo-peptide). This invention also relates to nucleic acid sequence construct encoding such chimeric molecule or encoding portions of these chimeric molecules.

Background

It is currently agreed that mitochondria play an important role in controlling life and death of cells (apoptosis; Kroemer and Reed 2000, Nature Medicine). It appears both that an increasing number of molecules involved in the transduction of the signal and also many metabolites and certain viral effectors act on mitochondria and influence the permeabilisation of mitochondrial membranes. Using mitochondrial-specific pro-apoptotic agent would seem to be an emerging concept in cancer therapy (Costantini et al 2000, Journal of the National Cancer Institute). Similarly, it might be possible to use cytoprotective molecules, thanks to their ability to stabilize mitochondrial membranes, in the treatment of illnesses where there is excessive apoptosis (neurodegenerative diseases, ischemia, AIDS, fulminant hepatitis, etc.).

Mitochondrial membrane permeabilisation (MMP) is a key event of apoptotic cell death associated with the release of caspase activators and caspase-independent death effectors from the intermembrane space, dissipation of the inner transmembrane potential ($\Delta\Psi_m$), as well as a perturbation of oxidative phosphorylation (Green and Reed, 1998; Gross *et al.*, 1999; Kroemer and Reed, 2000; Kroemer *et al.*, 1997; Lemasters *et al.*, 1998; Vander Heiden and Thompson, 1999; Wallace, 1999). Pro- and anti-apoptotic members of the Bcl-2 family regulate inner and outer MMP through interactions with the adenine nucleotide translocation (ANT; in the inner membrane, IM), the voltage-dependent anion channel (VDAC; in the outer membrane, OM), and/or through autonomous channel-forming activities (Desagher *et al.*, 1999; Gross *et al.*, 1999; Kroemer and Reed, 2000; Marzo *et al.*, 1998; Shimizu *et al.*, 1999; Vander Heiden and Thompson, 1999). ANT and VDAC are major components of the permeability transition pore complex (PTPC), a polyprotein structure organized at sites at which the two mitochondrial membranes are apposed (Crompton, 1999; Kroemer and Reed, 2000).

The mitochondrial phase is under the control of Bcl-2 family of oncogenes and anti-oncogenes (for review: 5; 28) involved in more than 50% of cancers (29). All members of Bcl-2 family play an active role in the regulation of apoptosis, some of them being proapoptotic (Bax, Bak, Bcl-X_S, Bad, etc.) and others, being antiapoptotic (Bcl-2, Bcl-X_L, Bcl-w, Mcl-1, etc.) (G. Kroemer, *Nat Med* 3, 614-20 (1997)).

The mitochondrial megachannel is a polyprotein complex formed in the contact site between the inner and the outer mitochondrial membranes that participate in the regulation of mitochondrial membrane permeability. It is composed of a set of proteins including mitochondrion-associated hexokinase (HK), porin (voltage-dependent anion channel or VDAC), adenine nucleotide translocation (ANT), peripheral benzodiazepin receptor (PBR), creatine kinase (CK), and cyclophilin D, as well as Bcl-2 family members. In physiological conditions, PTPC controls the mitochondrial calcium homeostasis via the regulation of its conductance by the mitochondrial pH, the $\Delta\Psi_m$, NAD/NAD(P)H redox equilibrium and matrix protein thiol oxidation. (M. Zoratti, I. Szabo, *Biochim, Biophys Acta* 1241, 139-76 (1995). S. Shimizu, M. Narita, Y. Tsujimoto, *Nature* 399, 483-487 (1999). M. Crompton, *Biochem J* 341, 233-249 (1999). K. Woodfield, A. Ruck, D. Brdiczka, A. P. Halestrap, *Biochem J* 336, 287-90 (1998).

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F. Ichas, L. Jouaville, J. Mazat, *Cell* 89, 1145-53 (1997)).

Apoptosis and related forms of controlled cell death are involved in a great number of illness. Excess or insufficiency of cell death processes are involved in auto-immune and neurodegenerative diseases, cancers, ischemia, and pathological infections or diseases such as viral and bacterial infections. Just few examples illustrating the virtually ubiquitous involvement of mitochondria in diseases associated with the abnormal control of cell death will be mentioned here.

In different models of ischemia (heart, liver, kidney or brain), using molecules that are capable of stabilising mitochondrial membranes, such as CsA for example (or its analogous non-immunosuppressor –Me-Val4-CsA) has made it possible to reduce massive apoptosis and its acute consequences at the level of the organ. In addition, VDAC is indispensable for the destruction of neurons of the rat hippocampus after hypoxic reperfusion. In the area of neurodegenerative diseases, a great many observations suggest close links with mitochondrial control of apoptosis (see Kroemer and Reed 2000, *Nature Medicine*). The neurotoxin –methyl-4-phenylpyridinium induces mitochondrial permeability transition and the exit of cytochrome c. Poisoning by mitochondrial toxins such as nitro-propionic acid or rotenone provokes in primates and rodents a Huntington-disease type of illness.

PTPC is a dynamic protein complex located at the contact site between the two mitochondrial membranes, its opening allowing the free diffusion of solutes < 1500 Da on the inner membrane. Formation of PTPC involves the association of proteins from different compartments, hexokinase (cytosol), porin, also called voltage-dependent anion channel (VDAC, outer membrane), peripheral benzodiazepin receptor (PBR, outer membrane), ANT (inner membrane) and cyclophilin D (matrix). PTPC has been implicated in many examples of apoptosis due to its capacity to integrate multiple pro-apoptotic signal transduction pathways and due to its control by proteins from Bcl-2/Bax family. The Bcl-2 family comprises death inhibitory (Bcl-2-like) and death inducing (Bax-like) members which respectively prevent or facilitate PTPC opening. Bax and Bcl-2 reportedly interact with VDAC and ANT within PTPC. In physiological conditions, ANT is a specific antiporter for ADP and ATP. However, ANT can also form a lethal pore upon interaction with different pro-apoptotic agents. including Ca^{2+} ,

atractyloside, HIV-1 Vpr-derived peptides and pro-oxidants. Mitochondrial membrane permeabilization may also be regulated by the non-specific VDAC pore modulated by Bcl-2/Bax-like proteins in the outer membrane (12; 16). and/or by changes in the metabolic ATP/ADP gradient between the mitochondrial matrix and the cytoplasm (17).

There is a need in the art for cytoprotective molecules in ischemia, neurodegenerative diseases, fulminant hepatitis and viral infections.

Another application of the chimeric molecule according the invention can be contemplated for the preparation of cosmetics or for preventing early death of plants or vegetables or flowers particularly for preventing the opening of the PTPC.

Conventional chemotherapeutic agents are limited in their therapeutic effectiveness by severe side effects due to their poor selectivity for tumors. The development of monoclonal antibodies (and ScFv) against specific tumor antigens and the identification of homing peptides specific for tumor vascularisation have made it possible to consider enhancing the selectivity of anticancer drugs by a targeted delivery approach. However, such reported attempts using monoclonal antibodies and the anticancer drugs doxorubicin (Trail P.A., et al 1993 Science 261:212), metotrexate (Kanellos J. et al., 1985 J Natl Cancer Inst 75:319), and Vinca alkaloids (Starling J.J. et al., 1991 Cancer Res 41:2965), have been largely unsuccessful. These antibody-drug conjugates were only moderately potent and usually less cytotoxic than the corresponding unconjugated drugs. In fact, antigen-specific cytotoxicity toward cultured tumor cells was rarely demonstrated. *In vivo* therapeutic effects with these conjugates in tumor xenograft animal models were in general observed only when the treatments were commenced before the tumors were well established or when exceedingly large doses (up to 90 mg/kg, drug equivalent dose) were used. It is, therefore, not surprising that in human clinical trials, no significant antitumor effects were observed with these agents (Elias D.J. et al., 1994 Am Respir Crit Care Med 150:1114) (Schneck D. et al., 1990). Indeed, the peak circulating serum concentrations of conjugates were only in the same range as their *in vitro* IC50 value and thus, capable of eliminating at best only about 50% of tumor cells.

These observations led to the conclusion that the previous attempts at delivering therapeutic doses of cytotoxic drugs via monoclonal antibodies have met with little success in clinical trials because of inappropriate choice of drug. One possible (partial-) solution was to

conclude that immunoconjugates must be composed of drugs possessing much higher potency than the clinically used anticancer agents if therapeutic levels of conjugate at the tumor sites are to be achieved in patients. Effectively, such toxins, including maytansinoides, enediynes, or intercalating agents CC1065, were shown to be 100 to 1000-fold more cytotoxic than the chemotherapeutic agents doxorubicin, methotrexate, and Vinca alkaloids (Chari RVJ et al., 1995 Cancer Res 55:4079) (Chari RVJ et al., 1992, Cancer Res 52:127).

Another approach termed "Adept" was also designed. This antibody-directed enzyme prodrug therapy (Adept) is based upon the use of a monoclonal antibody to target an enzyme at the tumor cell surface, which ultimately is expected to selectively deliver an antitumor drug from a suitable inactive prodrug. In both cases, clinical trials are in progress; however, since today none of them have been introduced in cancer chemotherapy, there is a need for new tools to kill target tumor cells. Bagshawe KD, 1990. Antibody-directed enzyme/prodrug therapy (ADEPT). Biochem Soc Trans. 18(5):750-2. Melton RG, Sherwood RF. 1996 Antibody-enzyme conjugates for cancer therapy. J Natl Cancer Inst, 88(3-4):153-65. Rihova B. 1997; Targeting of drugs to cell surface receptors. Crit Rev Biotechnol. 17(2):149-69. Hudson PJ. 2000. Recombinant antibodies: a novel approach to cancer diagnosis and therapy. Expert Opin Investig Drugs 9(6):1231-42.

Recently, the mitochondrion has been proposed as a novel prospective target for chemotherapy-induced apoptosis (1-7). Indeed, four different anti-cancer agents, including the resinoid acid-derivative CD437, lonidamine, betulinic acid, and arsenite, have been shown to induce cancer cell apoptosis by a direct action on mitochondria. The interaction of these anti-cancer agents with mitochondria results in an increase of the permeability of the inner mitochondrial membrane due, at least in part, to the opening of the permeability transition pore complex (PTPC). PTPC opening leads to swelling of the mitochondria matrix, the dissipation of the inner transmembrane potential ($\Delta\Psi_m$), enhanced generation of reactive oxygen species (ROS), and the release of apoptogenic proteins from the intermembrane space to the cytoplasm. Such mitochondrial apoptogenic effectors include the caspase activator cytochrome c, apoptosis inducing factor (AIF), and pro-caspases (2-6). All the signs of apoptosis induced by CD437, lonidamine, betulinic acid, and arsenite are prevented by two agents acting on specific PTPC proteins, namely cyclosporin A (CsA, a cyclophilin D ligand) and bongkrekic acid (BA, a

ligand of the adenine nucleotide translocase (ANT)). It thus appears that PTPC opening is a critical event of apoptosis triggered by these agents.

Mastoparan, a peptide isolated from wasp venom, is the first peptide known to induce mitochondrial membrane permeabilization via a CsA-inhibitable mechanism and to induce apoptosis via a mitochondrial effect when added to intact cells. This peptide has an α -helical structure and possesses some positive charges that are distributed on one side of the helix. A similar peptide (KLAKLAKKLAKLAK or (KLAKLAK)₂ (K = lysine, L = alanine, and A = leucine) has been found recently to disrupt mitochondrial membranes when it is added to purified mitochondria, although the mechanisms of this effect have not been elucidated.

The vasculature of individual tissues is highly specialized. The endothelium in lymphoid tissues expresses tissue-specific receptors for lymphocyte homing, and recent work utilizing phage homing has revealed an unprecedented degree of specialization in the vasculature of other normal tissues. *In vivo* screening of libraries of phage that displace random peptide sequences on their surfaces has yielded specific homing peptides for a large number of normal tissues. The tissue-specific endothelial molecules to which the phage peptides home may serve as receptors for metastasizing malignant cells. Probing of tumor vasculature has yielded peptides that home to endothelial receptors expressed selectively in angiogenic neovasculature. These receptors, and those specific for the vasculature of individual normal tissues, are likely to be useful in targeting therapies to specific sites. Ruoslahti E, Rajotte D. 2000; An address system in the vasculature of normal tissues and tumors. *Annu Rev Immunol*. 18:813-27.

Ellerby et al. recently have fused the mitochondriotoxic (KLAKLAK)₂ motif to a targeting peptide that interacts with endothelial cells. Such a fusion peptide is internalized and induces mitochondrial membrane permeabilization in angiogenic endothelial cells and kills MDA-MD-435 breast cancer xenografts transplanted into nude mice. Similarly, a recombinant chimeric protein containing interleukin 2 (IL-2) protein fused to Bax selectively binds to and kills IL-2 receptor-bearing cells *in vitro*. Thus, specific cytotoxic agents that target surface receptors, translocate into the cytoplasm, and induce apoptosis via mitochondrial membrane permeabilization might be useful in treating cancer.

There is a need in the art for the selective eradication of transformed cells. One strategy is to target a toxic agent to selected cell types. More particularly, there exists a need in the art for method and reagents for regulating mitochondrial permeabilization and apoptosis.

Summary of the Invention

In order to overcome at least some of the limitations of the prior art, the present invention provides a peptidic or pseudo-peptidic family of polyfunctional molecules containing a cell-targeting part (termed TARG), a PTPC-interacting part (termed TOX/SAVE), and a facultative mitochondrial localisation sequence (MLS). In a preferred embodiment of the invention, the TOX/SAVE portion of the said polyfunctional molecule is a peptide or peptidomimetic molecule which interact directly with the Adenine Nucleotide Translocator (ANT) a central component of the PTPC

Thus, the present invention includes two categories of targeted cell death regulatory molecules:

- TARG-(MLS)-TOX is a polyfunctional molecule which induces a PTPC-dependent mitochondrial membrane permeabilisation and consequent cell death.
- TARG-(MLS)-SAVE is a polyfunctional molecule which protects cells from mitochondrial membrane permeabilisation and consequently from cell death through interaction with the PTPC and/or ANT.

The invention further provides a vector encoding a chimeric polypeptide of the invention. Also, the invention provides a recombinant host cell comprising a vector of the invention.

Further, the invention provides a cancer cell having a tumor-associated antigen on the surface thereof to which the chimeric polypeptide of the invention is bound via the antibody or antibody fragment of the chimeric polypeptide. The invention also provides methods for detecting cancer cells.

The invention also provides methods for inducing or preventing apoptosis with polypeptides of the invention. The invention provides methods for inducing apoptosis in tumor cells. The invention provides methods for inducing apoptosis in virus infected cells.

The invention further provides hybridomas producing polypeptides of the invention. The invention also provides monoclonal antibodies produced by these hybridomas.

The invention also provides methods for identifying active agents of interest that interact with the PTPC. The invention also provides methods for identifying active agents of interest that interact with ANT peptide. The invention also provides methods for identifying mitochondrial antigens.

The invention also provides methods of treatment or prevention of a pathological infection or disease by administering a polypeptide of the invention to a patient. The invention also provides pharmaceutical compositions comprising a polypeptide of the invention.

Brief Description of the Drawings

Figure 1 shows the nucleotide sequence of vector pACgp67-ScFv461.

Figure 2 shows the nucleotide sequence of vector pACgp67-ScFv350.

Figure 3 shows the nucleotide sequence of Vh and VL, from the clone therap 99B3.

Figure 4 shows the nucleotide sequence of Vh and VL from the clone therap.88E10.

Figure 5 shows the nucleotide sequence of Vh and VL from the clone therap.152C3.

Figure 6, 7, 8, 9, 10, 11 show surface plasmon resonance curves.

Figures 12 and 13 show the strategy for obtaining the ScFv-transfert vector.

Detailed Description of the Invention

It was recently discovered that the proapoptotic HIV-1-encoded protein Vpr induces mitochondrial membrane permeabilization via its physical and functional interaction with the mitochondrial inner membrane protein ANT (adenine nucleotide translocation, also called ADP/ATP carrier). This was shown using a variety of different techniques: surface plasmon resonance, electrophysiology, synthetic proteoliposomes, studies on purified mitochondria (respirometry, electron microscopy, organellofluorometry), as well as microinjection of intact cells. These discoveries are described in detail in U.S. Provisional Application No. 60/231,539 filed September 11, 2000, the entire disclosure of which is relied upon and incorporated by reference herein.

The present invention pertains to novel cytotoxic conjugates based on the association between a peptidic molecule (named pTox) interacting with the mitochondrial permeability transition pore complex (PTPC) and a molecule (named pTarg) able to target cells. The present

invention also pertains to novel cytoprotective conjugates based on the association between a peptidic molecule (named SAVE) interacting with the mitochondrial permeability transition pore complex (PTPC) and a molecule (named pTarg) able to target the cells to rescue. In a specific embodiment of this invention, a cytotoxic conjugate of the invention includes a viral derived pro-apoptotic peptide.

In one embodiment of the invention, the polyfunctional molecule TARG-(MLS)-TOX is a tumor specific molecule that selectively interact with a tumor cell or a specific mammalian cell type, where the polyfunctional molecule is selectively internalised by the mammalian or tumoral cell type, where the polyfunctional molecule interact with the PTPC and/or ANT and exhibits thereto a strong mitochondrio-toxicity leading to apoptosis or any cell death process.

In one embodiment of the invention, the polyfunctional molecule TARG-(MLS)-TOX exhibits a selective toxicity against angiogenic endothelial cells. In another embodiment of the invention, the polyfunctional molecule TARG-(MLS)-TOX exhibits a selective toxicity against tumor cells.

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is an antibody or a recombinant antibody fragment. In another embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is tumor homing peptide (example; CNGRC peptide; lung-homing peptide CGFECVRQCPERC).

In one embodiment of the invention, the TOX part of the polyfunctional molecule TARG-(MLS)-TOX is a peptide or a peptido-mimetic derived from the C-terminal part (amino-acids 52 to 96) of the HIV-1 Vpr protein.

In one embodiment of the invention, the TOX part of the polyfunctional molecule TARG-(MLS)-TOX is a pro-apoptotic Bcl-2 family member such as the Bax or Bid proteins, or a fragment thereof.

In one embodiment of the invention, the TOX part of the polyfunctional molecule TARG-(MLS)-TOX is a D-peptide, is a Ψ -peptide or a retro-inverso peptide chosen among the group of peptidic sequences described in table 1:

Table 1:

Name	TOX Peptidic Sequences
Vpr71-82	HFRIGCRHSRIG

Vpr71-82[R73,77,80K]	HFKIGCKHSGKIG
Vpr71-96	HFRIGCRHSRIGIIQRRTRNGASKS
Vpr71-96[R73,77,80K]	HFKIGCKHSGKIGIIQRRTRNGASKS
Vpr52-96	DTWTGVEALIRILQQLFIHFRIGCRHSRIGIIQRRTRNGASKS
Vpr52-96[R73,77,80K]	DTWTGVEALIRILQQLFIHFKIGCKHSGKIGIIQRRTRNGASKS
Vpr52-96[L60,67A]	DTWTGVEAAIRILQQALFIHFRIGCRHSRIGIIQRRTRNGASKS
Vpr52-82	DTWTGVEALIRILQQLFIHFRIGCRHSRIG
Vpr52-82[R73,77,80K]	DTWTGVEALIRILQQLFIHFKIGCKHSGKIG
Histatin5 Candida Albicans	DSHARKRHHGYKRKFHEKHSHRGY
Mastoparan Vespula Lewisii	INLKALAALAKKIL
hNUR77(555-568)	LSRLLGKLPELRTL
hNTR(368-381) neutrotrophin receptor	ATLDALLAALRRIQ
Bid(84-100)	RNIARHLAQVGDSMRDR
Bax(57-72)	KKLSECLKRIGDELDS
Bax(72-87)	GQVGRQLAIIIGDDINR
HBX(70-78)	ALRFTSARR
DCC(1376-1390)	KTHVKTASLGLAGKA
ANT ₁ (104-116)	DRHKQFWRYFAGN
ANT ₂ (104-116)	DKRTQFWRYFAGN
ANT ₃ (104-116)	DKHTQFWRYFAGN
ANT ₁ (104-116 [A114P])	DRHKQFWRYFPGN
ANT ₂ (104-116)[A114P]	DKRTQFWRYFPGN
ANT ₃ (104-116)[A114P]	DKHTQFWRYFPGN
ANT _{1,2,3} (117-134)	LASGGAAGATSLCFVYPL
ANT ₁ (104-134)	DRHKQFWRYFAGNLASGGAAGATSLCFVYPL
ANT ₂ (104-134)	DKRTQFWRYFAGNLASGGAAGATSLCFVYPL
ANT ₃ (104-134)	DKHTQFWRYFAGNLASGGAAGATSLCFVYPL
ANT ₁ (104-134)[A114P]	DRHKQFWRYFPGNLASGGAAGATSLCFVYPL
ANT ₂ (104-134 [A114P])	DKRTQFWRYFPGNLASGGAAGATSLCFVYPL
ANT ₃ (104-134) [A114P]	DKHTQFWRYFPGNLASGGAAGATSLCFVYPL
Vpr 52-96 [C76S]	DTWTGVEALIRILQQLFIHFRIGSRHSRIGIIQRRTRNGASKS
HTLV-1p13II	¹⁹ PSLRVWRLCARRLV ₃₂

Bad103-127	NLWAAQRYGRELRRMSDEFVDSFKK
Bax52-76	QDASTKKLSECLKRIGDELDSNMEL

In one embodiment of the invention, the SAVE part of the polyfunctional molecule TARG-(MLS)-SAVE is a L-peptide, a D-peptide or a retro-inverso peptide chosen among the group of peptidic sequences described in table II:

Name	SAVE Peptidic Sequences
ANT ₁ (104-116)	DRHKQFWRYFAGN
ANT ₂ (104-116)	DKRTQFWRYFAGN
ANT ₃ (104-116)	DKHTQFWRYFAGN
ANT _{1,2,3} (117-134)	LASGGAAGATSLCFVYPL
ANT ₁ (104-134)	DRHKQFWRYFAGNLASGGAAGATSLCFVYPL
ANT ₂ (104-134)	DKRTQFWRYFAGNLASGGAAGATSLCFVYPL
ANT ₃ (104-134)	DKHTQFWRYFAGNLASGGAAGATSLCFVYPL

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MIS)-SAVE is a L-peptide, a D-peptide or a retro-inverso peptide chosen among the group of peptidic sequences described in table III:

ANTENNAPEDIA third helix (residues 43-58)	RQIKITFQNRMRMKTCK
HIV-1 Vpr 83-96 transduction domain	IIQQRRTNRNGASKS
HIV-1 Tat48-59 transduction domain	GRKKRRQRRPP
HIV-1 Tat49-57 transduction domain	RKKRRQRRR
pep-1	KETWWETWWTEW

In one embodiment of the invention, the Targ part of the polyfunctional molecule TARG-(MLS)-TOX is the decanoic acid $\text{CH}_3(\text{CH}_2)_8\text{CO}-$.

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is an antibody, a recombinant antibody, a recombinant antibody fragment or a ScFv (single chain fragment variable).

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is encoded by the following vector pACgp67-ScFv461 (figure 1).

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is encoded by the following vector pACgp67-ScFv350 (figure 2).

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX is a tumor homing peptide as defined by Ellerby et al in PCT/US00/01602.

In one embodiment of the invention, the TARG part of the polyfunctional molecule TARG-(MLS)-TOX/SAVE is a brain or kidney homing peptide as defined by Pasqualini R, Ruoslahti (in Nature 1996 Mar 28;380(6572):364-6. Organ targeting in vivo using phage display peptide libraries).

In one embodiment of the invention, pTox is the Vpr peptide of HIV-1 or a fragment thereof. Protein R (Vpr) of human immunodeficiency virus type 1 (HIV-1) is a virion-associated viral gene product with an average length of 96 amino acids, and a molecular weight of approximately 15 kD. Vpr is a highly conserved viral protein among HIV, simian immunodeficiency viruses (SIV). See Yuqi Zhao and Robert T. Elder, "Yeast Perspectives on HIV-1 VPR," Frontiers in Bioscience 5, d905-916, December 1, 2000.

Vpr has been characterized as an oligomer, and is thought to be divided into three domains on the basis of its structural features: an amino-terminal, negatively charged region that is predicted to form an amphipathic α helix (amino acids 17 to 34); a central hydrophobic domain (amino acids 35 to 75); and a carboxy-terminal, positively charged domain (amino acids 80 to 96). Mutational analysis of Vpr suggests that the nuclear import, virion incorporation, and cell cycle arrest of Vpr are mediated by the distinct functional domains. A structural motif within an amino-terminal helix appears to be important for packaging of Vpr into virions and for maintaining the stability of the protein. A central hydrophobic region, especially the leucine-isoleucine (LR) domain, is reported to be involved in the nuclear localization of Vpr. The cell

cycle arrest function of Vpr was found to be largely located within a carboxy-terminal, positively charged region. *See* Tomoyuki Yamaguchi, Nobumoto Watanabe, Hiromitsu Nakauchi, and Atsushi Koito, "Human Immunodeficiency virus type 1 Vpr Modifies Cell Proliferation via Multiple Pathways," *Microbiol, Immunol.*, 43(5), 437-447, 1999.

The amino acid sequence of human immunodeficiency virus type 1 viral protein R (Vpr) is shown below:

MEQAPEDQGPQREPYNEWTLLEELKSEAVRHFPRIWLHNLGQHIYE
TYGDTWAGVEAIIRILQQLLFHFRIGCRHSRIGVTRQRRARNGASRS.

Vpr and peptides containing conserved H(F/S)RIG repeat motifs can rapidly penetrate human CD4 cells, and cause mitochondrial dysfunction and death by apoptosis. More particularly, recombinant Vpr and C-terminal peptides of Vpr containing the conserved sequence HFRIGCRHSRIG can cause permeabilization of CD4⁺ T lymphocytes, a dramatic reduction of mitochondrial membrane potential, and finally cell death. Vpr and Vpr peptides containing the conserved sequence rapidly penetrate cells, co-localize with the DNA, and cause increased granularity and formation of dense apoptotic bodies. Vpr treated cells undergo apoptosis, and this was confirmed by demonstration of DNA fragmentation. *See* C. Arunagiri, I. Macreadie, D. Hewish and A. Azad, "A C-terminal domain of HIV-1 accessory protein Vpr is involved in penetration, mitochondrial dysfunction and apoptosis of human CD4⁺ lymphocytes," *Apoptosis* 1997; 2: 69-76.

Using a yeast model system, it has been confirmed that there is a cytotoxic activity associated with the C-terminal portion of Vpr, particularly the sequence HFRIGCRHSRIG. Vpr and portions of Vpr containing the sequence HFRIGCRHSRIG can kill a range of mammalian cells including human lymphocytes. *See* I.G. Macreadie, A. Kirkpatrick, P.M. Strike, and A.A. Azad, "Cytotoxic Activities of HIV-1 VPR and Sac1p peptides Bioassayed in Yeast," *Protein and Peptide Letters*, Vol. 4, No. 3, pp. 181-186, 1997.

The C-terminal moiety (Vpr52-96), within an α -helical motif of 12 amino acids (Vpr71-82), contain several critical arginine (R) residues (R73, R77, R80), which are strongly conserved among different pathogenic HIV-1 isolates. L.G. Macreadie, et al., *Proc. Natl. Acad. Sci. USA* 92, 2770-2774 (1995). I.G. Macreadie, et al., *FEBS Lett.* 410, 145-149 (1997). E. Jacotot, et al., *J. Exp. Med.* 191, 33-45 (2000). Thus, the pro-apoptotic portion (pTox) of the chimeric

polypeptide of the invention can contain, for example, the sequence HFRIGCRHSRIG (HIV-1 Vpr⁷¹⁻⁸²), HFKIGCKHSKIG, Vpr⁷¹⁻⁹⁶, Vpr⁵²⁻⁹⁶, or a pseudo peptidic variant such as D[HFRIGCRHSRIG].

Other variants of Vpr peptides can also be employed in this invention. Peptide fragments of Vpr encompassing a pair of H(F/S)RIG sequence motifs (residues 71-75 and 78-82 of HIV-1 Vpr) have been shown cause cell membrane permeabilization and death in yeast and mammalian cells. Peptide Vpr⁵⁹⁻⁸⁶ (residues 59-86 of Vpr) forms an α -helix encompassing residues 60-77, with a kink in the vicinity of residue 62. It has been shown that the first of the repeated sequence motifs (HFRIG) participates in a well-defined α -helical domain, whereas the second (HSRIG) lay outside the helical domain and forms a reverse turn followed by a less ordered region. On the other hand, peptides Vpr⁷¹⁻⁸² and Vpr⁷¹⁻⁹⁶, in which the sequence motifs are located at the N-terminus, were largely unstructured under similar conditions, as judged by their C²H chemical shifts. Thus, it has been shown that the HFRIG and HSRIG motifs adopt α -helical and turn structures, respectively, when preceded by a helical structure, but are largely unstructured in isolation. There are implications of these findings for interpretation of the structure-function relationships of synthetic peptides containing these motifs. For example, since the HFRIG and HSRIG sequence motifs adopt helical and turn structures, respectively, when preceded by a helical structure, as in full-length Vpr, but are largely unstructured in isolation, 7-8 residues, sufficient to support at least 1-2 turns of helix, should be included at the N-terminus of Vpr when used as the pTox component of the chimeric polypeptides of the invention to ensure that they are able to adopt the same structure as in the full-length protein. See Shenggen Yao, Allan M. Torres, Ahmed A. Azad, Ian G. Macreadie and Raymond S. Norton, "Solution Structure of Peptides from HIV-1 Vpr Protein that Cause Membrane Permeabilization and Growth Arrest," J. Peptide Sci. 4: 426-435 (1998). While the Vpr gene codes for a protein of 96-amino-acids, variations have been observed, e.g., Vprs from HIV-1_{HXB2} have 97 and 90-amino-acid residues, respectively. It will be understood that these variants can also be employed in this invention.

For the most effective toxicity, HFRIGCRHSRIG should be surrounded on each side by about eight amino acids from the native sequence. Vpr polypeptides and peptides of greater than 9 amino acids that inhibit or augment Vpr binding, mitochondrial membrane permeabilization, or apoptosis can also be employed in the invention, as well as peptides that are at least 10-20, 20-

30, 30-50, 50-100, and 100-365 amino acids in size. DNA fragments encoding these polypeptides and peptides are encompassed by the invention. Flanking residues should not disrupt the helical structures described above.

The Vpr variants and other viral apoptotic peptides can be assessed for their ability to mediate apoptosis, and thus their suitability for use as pTox in the invention. It is understood that many techniques could be used to assess binding of Vpr or another viral apoptotic peptide to ANT, and that these embodiments in no way limit the scope of the invention. For example, in one embodiment, surface plasmon resonance is used to assess binding of Vpr or another viral apoptotic peptide to ANT. In another embodiment, electrophysiology is used to assess binding of Vpr or another viral apoptotic peptide to ANT. In another embodiment, purified mitochondria are used to assess binding of Vpr or another viral apoptotic peptide to ANT. In another embodiment, synthetic proteoliposomes are used to assess binding of Vpr or another viral apoptotic peptide to ANT. In another embodiment, microinjection of live cells is used to assess binding of Vpr or another viral apoptotic peptide to ANT. These techniques are described in U.S. Provisional Application No. 60/231,539.

In another embodiment, the yeast two-hybrid system developed at SUNY (described in U.S. Patent No. 5,282,173 to Fields et al.; J. Luban and S. Goff., *Curr Opin. Biotechnol.* 6:59-64, 1995; R. Brachmann and J. Boeke, *Curr Opin. Biotechnol.* 8:561-568, 1997; R. Brent and R. Finley, *Ann. Rev. Genet.* 31:663-704, 1997; P. Bartel and S. Fields, *Methods Enzymol.* 254:241-263, 1995) can be used to screen for Vpr-ANT interaction as follows. Vpr, or portions thereof, or another viral apoptotic peptide, responsible for interaction, can be fused to the Gal4 DNA binding domain and introduced, together with an ANT molecule fused to the GAL 4 transcriptional activation domain, into a strain that depends on GAL4 activity for growth on plates lacking histidine. Interaction of the Vpr polypeptide or another viral apoptotic peptide with an ANT molecule allows growth of the yeast containing both molecules and allows screening for the molecules that inhibit or alter this interaction (i.e., by inhibiting or augmenting growth). In an alternative embodiment, a detectable marker (e.g. β -galactosidase) can be used to measure binding in a yeast two-hybrid assay.

Alternatively, the binding properties of Vpr peptide fragments or another viral apoptotic peptide can be determined by analyzing the binding of Vpr peptide fragments or another viral

apoptotic peptide to ANT-expressing cells by FACS analysis. This allows the characterization of the binding of the peptides, and the discrimination of relative abilities of the peptide to bind to ANT. *In vitro* binding assays with Vpr or another viral apoptotic peptide can similarly be used to characterize ANT binding activity.

In another specific embodiment, a cytotoxic conjugate of the invention includes an adenine nucleotide translocation (ANT)-derived pro-apoptotic peptide. The pro-apoptotic portion (pTox) of the conjugate can contain, for example, the sequence DKRTQFWRYFPGN (hANT₂104-116[A114P]) or a pseudo-peptidic variant such as [DKRTQFWRYFPGN].

In another specific embodiment, a cytoprotective conjugate of the invention includes ANT-derived anti-apoptotic peptides. The anti-apoptotic portion (pSave) of the conjugate can contain, for example, the sequence DKRTQFWRYFAGN (hANT₂104-116), the sequence LASGGAAGATSLCFVYPL (ANT 117-134) or a pseudo-peptidic variant such as D[DKRTQFWRYFPGN].

The pTarg component of the chimeric polypeptide of the invention can be an antibody or an antibody fragment. The antibody or antibody fragment can be all or part of a polyclonal or monoclonal antibody. The term "antibodies" is meant to include polyclonal antibodies, monoclonal antibodies, fragments thereof, as well as any recombinantly produced binding partners. Antibodies are defined to be specifically binding if they bind with a K_a or greater than or equal to about 10^7 M^{-1} . Affinities of binding partners or antibodies can be readily determined using conventional techniques, for example those described by Scatchard *et al.*, *Ann. N.Y. Acad. Sci.*, 51:660 (1949).

As used herein, the term "antibody fragment" includes the following:

Fc	A constant region dimer lacking C _H 1
Fab	A light chain dimerized to V _H -C _H 1 resulting from papain cleavage; this is monomeric since papain cuts above the hinge cystines
F(ab) ₂	A dimer of Fab' resulting from pepsin cleavage below the hinge disulfides; this is bivalent and can precipitate antigen

Fab'	A monomer resulting from mild reduction of F(ab)' ₂ : an Fab with part of the hinge
Fd	The heavy chain portion of Fab (V _H -C _{H1}) obtained following reductive denaturation of Fab
Fv	The variable part of Fab: a V _H -V _L dimer
Fb	The constant part of Fab: a C _{H1} -C _L dimer
pFc'	A C _{H3} dimer

Fragments of monoclonal antibodies are of particular interest as small antigen targeting molecules. Antibody fragments are also useful for the assembly of the chimeric polypeptides of the invention designed to carry other pTox agents, such as a therapeutic conjugate. For *in vivo* applications, fragments of antibodies are of interest due to their altered pharmacokinetic behavior, which is useful for cancer therapy with cytotoxic agents, and for their rapid penetration into body tissues, which offer advantages for therapy techniques.

An antibody fragment of particular interest for use in the invention is a minimal Fv fragment with antigen-binding activity. The two chains of the Fv fragment are less stably associated than the Fd and light chain of the Fab fragment with no covalent bond and less non-covalent interaction, but nevertheless functional Fv fragments have been expressed for a number of different antibodies. Two strategies can be employed to stabilize the Fv fragments used in the invention: firstly, mutating a selected residue on each of the V_H and V_L chains to a cysteine to allow formation of a disulphide bond between the two domains; and secondly, the introduction of a peptide linker between the C-terminus of one domain and the N-terminus of the other, such that the Fv is produced as a single polypeptide chain known as a single-chain Fv.

Thus, single-chain Fvs (ScFvs), recombinant V_L and V_H fragments covalently tethered together by a polypeptide link and forming one polypeptide chain, are useful in this invention. For expression of Fv genes, several systems can be effectively used, including myeloma cells, insect, yeast, and *Escherichia coli* cells. Expression in *E. coli* has been a frequently used production method, with both intracellular expression and secretion enabling high yields of ScFv to be made.

The production of ScFv molecules requires the identification of a suitable peptide linker to span the 35-40 Å distance between the C-terminus of one domain and the N-terminus of the other and allow correct folding and assembly of the Fv structure. Several different types of linkers have been used and shown to result in functional ScFv. Polypeptides with the average length of 3-18 amino acids are usually used as links. They can be rich in serine and/or glycine residues, which introduce flexibility, or in charged glutamic acid and/or lysine residues, which improve solubility. Linkers can be selected from searching existing protein structures for protein fragments of the appropriate length and conformation, or by designing them *de novo* based on simple, flexible structures, such as the 15 amino acid sequence (Gly₄Ser)₃.

Active single-chain Fv molecules in both of the two possible orientations, V_H-linker-V_L or V_L-linker-V_H are useful in the invention; however, for some antibodies one particular orientation may be preferable as a free N-terminus of one domain, or C-terminus of the other, may be required to retain the native conformation and thus full antigen binding.

The ScFv may be susceptible to aggregation, with dimers, trimers, and multimers formed. The potential of forming dimers or other multimers with very short linkers, or no linker at all, can be exploited to produce stable pTarg structures. Such an approach can also be used to create pTarg molecules with two different binding specificities by fusing the V_H of an antibody of one specificity to the V_L of another and vice versa.

Fv's stabilized by disulphide linkages can also be employed as the pTarg component of the chimeric polypeptide of the invention. The introduction of a disulphide bond between the V_H and V_L domains to form a disulphide-linked Fv requires the identification of residues in close proximity on each chain, which are unlikely to affect directly the conformation of the binding site when mutated to cysteine, and will be capable of forming a disulphide bond without introducing strain into the structure of the Fv. Sites have been identified in both CDR regions and framework regions, which appear to result in the formation of such disulphide bonds and allow the production of stabilized Fv fragments which retain antigen-binding characteristics.

Due to small size, rapid clearance *in vivo*, stability, and easy engineering, ScFvs employed in this invention have various applications in the treatment of diseases, particularly of cancer. ScFvs can exhibit the same affinity and specificity for antigen as monoclonal antibodies. Dozens of ScFvs with different specificities have been constructed. They are useful for genetic

fusion to the potent toxins (pTox). If the monovalency of ScFv is a disadvantage, constructs with di- or multivalency with increased combining efficiency can be employed.

In a preferred embodiment of the invention, the targeting part (pTarg) of the cytotoxic conjugate is a recombinant portion (ScFv) of a tumor specific antibody, such as the ScFv versions of the M350 and V461 monoclonal antibodies. The hybridoma has been deposited at the CNCM on January 24, 2001, under the Accession Number I-2617.

The pTarg component of the chimeric polypeptide of the invention is preferably a monoclonal antibody or a fragment thereof. Monoclonal antibodies to human cell antigens are preferred. Many tumor-associated antigens are now known and characterized, and antibodies to these allow targeting to different tumor types. Useful tumor-associated antigens are absent on normal tissues and present at high levels on tumor cells, preferably homogeneously on all cells of the tumor. Antigen should also not be shed from the tumor into the blood.

Commonly used tumor-associated antigens and examples of antibodies raised against them are described in the following Table.

Antigen	Tumor type	Representative antibody
Tumor-associated glycoprotein 72 (TAG72), 72 kDa glycoprotein	Pancarcinoma	B72.3, CC49
Carcinoembryonic antigen (CEA), 180 kDa glycoprotein	Pancarcinoma	NP-4, A5B7
Polymorphic epithelial mucin (PEM), >100 kDa glycoprotein	Ovarian, breast, lung	HMFG1
Epithelial membrane antigen (EMA), 40 kDa glycoprotein	Colorectal (and other epithelial tumors)	17-1A
epidermal growth factor receptor (EGFR), 175 kDa glycoprotein	Breast, lung	425
p185 ^{HER2} /c-erb-B2		

Antigen	Tumor type	Representative antibody
(185 kDa glycoprotein)	Breast, lung	4D5
Prostate-specific membrane antigen (PSMA), 100 kDa glycoprotein	Prostrate	7E11-C5.3
CD33 67 kDa glycoprotein	Myeloid leukemia	P67.6,M195
CD 20 35 kDa glycoprotein	Lymphoma	C2B8
GD2 ganglioside	Melanoma, neuroblastoma	14-18

An important consideration is the absolute amount of antibody localized to the tumor site. Therefore, the ideal molecule would localize to the tumor in large amounts, delivering a high dose of pTox while clearing rapidly from the circulation and the rest of the body, minimizing non-specific toxicity. Intact antibodies typically circulate for a long period of time and accumulate high levels of activity at the tumor site, whereas antibody fragments clear more rapidly, sparing the dose to normal tissues.

The antibody fragments can also be prepared by phage-display technology. Phage display is a selection technique, according to which an antibody fragment (ScFv) is expressed on the surface of the filamentous phage fd. For this, the coding sequence of the antibody variable genes is fused with the gene that encoded the minor coat phage protein III (g3p) located at the end of the phage particle. The fused antibody fragments are displayed on the virion surface and particles with the fragments can be selected by adsorption on insolubilized antigen (panning). The selected particles are used after elution to reinfect bacterial cells. The repeated rounds of adsorption and infection lead to enrichment. Bacterial proteases can cleave the bond between the g3p protein and antibody fragments, which results in the production of soluble antibody fragments by infected bacterial cells. To release the soluble ScFvs, an excision of the g3p gene is made or an amber stop codon between the antibody gene and the g3p gene is engineered.

Immunoglobins and certain variants thereof are known and many have been prepared in recombinant cell culture. For example, see U.S. Patent 4,745,055; EP 256,654; Faulkner *et al.*, Nature 298:286 (1982); EP 120,694; EP 125,023; Morrison, J. Immun. 123:793 (1979); Köhler *et al.*, P.N.A.S. USA 77:2197 (1980); Raso *et al.*, Cancer Res. 41:2073 (1981); Morrison *et al.*, Ann. Rev. Immunol. 2:239 (1984); Morrison, Science 229:1202 (1985); Morrison *et al.*, P.N.A.S. USA 81:6851 (1984); EP 255,694; EP 266,663; and WO 88/03559. Reassorted immunoglobulin chains also are known. See for example U.S. patent 4,444,878; WO 88/03565; and EP 68,763 and references cited therein. DNA encoding immunoglobulin light or heavy chain constant regions is known or readily available from cDNA libraries or is synthesized. See for example, Adams *et al.*, Biochemistry 19:2711-2719 (1980); Gough *et al.*, Biochemistry 19:2702-2710 (1980); Dolby *et al.*, P.N.A.S. USA, 77:6027-6031 (1980); Rice *et al.*, P.N.A.S. USA 79:7862-7865 (1982); Falkner *et al.*, Nature 298:286-288 (1982); and Morrison *et al.*, Ann. Rev. Immunol. 2:239-256 (1984). These materials and techniques can be employed to synthesize the pTarg component of the chimeric polypeptide of the invention.

Polyclonal antibodies employed as the pTarg component of the chimeric polypeptide of the invention can be readily generated from a variety of sources, for example, horses, cows, goats, sheep, dogs, chickens, rabbits, mice, or rats, using procedures that are well known in the art. In general, purified cell surface proteins or glycoproteins or a peptide based on the amino acid sequence of cell surface proteins or glycoproteins that is appropriately conjugated is administered to the host animal typically through parenteral injection. The immunogenicity of cell surface proteins or glycoproteins can be enhanced through the use of an adjuvant, for example, Freund's complete or incomplete adjuvant. Following booster immunizations, small samples of serum are collected and tested for reactivity to cell surface proteins or glycoproteins. Examples of various assays useful for such determination include those described in *Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; as well as procedures, such as countercurrent immuno-electrophoresis (CIEP), radioimmunoassay, radio-immunoprecipitation, enzyme-linked immunosorbent assays (ELISA), dot blot assays, and sandwich assays. See U.S. Patent Nos. 4,376,110 and 4,486,530.

Monoclonal antibodies employed as the pTarg component can be readily prepared using well known procedures. See, for example, the procedures described in U.S. Patent Nos. RE

32,011, 4,902,614, 4,543,439, and 4,411,993; Monoclonal Antibodies, Hybridomas: *A New Dimension in Biological Analyses*, Plenum Press, Kennett, McKearn, and Bechtol (eds.), 1980. Briefly, the host animals, such as mice, are injected intraperitoneally at least once and preferably at least twice at about 3 week intervals with isolated and purified cell surface proteins or glycoproteins, conjugated cell surface proteins or glycoproteins, optionally in the presence of adjuvant. Mouse sera are then assayed by conventional dot blot technique or antibody capture (ABC) to determine which animal is best to fuse. Approximately two to three weeks later, the mice are given an intravenous boost of cell surface proteins or glycoproteins or conjugated cell surface proteins or glycoproteins. Mice are later sacrificed and spleen cells fused with commercially available myeloma cells, such as Ag8.653 (ATCC), following established protocols. Briefly, the myeloma cells are washed several times in media and fused to mouse spleen cells at a ratio of about three spleen cells to one myeloma cell. The fusing agent can be any suitable agent used in the art, for example, polyethylene glycol (PEG). Fusion is plated out in plates containing media that allows for the selective growth of the fused cells. The fused cells can then be allowed to grow for approximately eight days. Supernatants from resultant hybridomas are collected and added to a plate that is first coated with goat anti-mouse Ig. Following washes, a label, such as ¹²⁵I-labeled cell surface proteins or glycoproteins, is added to each well followed by incubation. Positive wells can be subsequently detected by autoradiography. Positive clones can be grown in bulk culture and supernatants are subsequently purified over a Protein A column (Pharmacia).

The monoclonal antibodies for the pTarg component can be produced using alternative techniques, such as those described by Alting-Mees *et al.*, "Monoclonal Antibody Expression Libraries: A Rapid Alternative to Hybridomas", *Strategies in Molecular Biology* 3:1-9 (1990), which is incorporated herein by reference. Similarly, binding partners can be constructed using recombinant DNA techniques to incorporate the variable regions of a gene that encodes a specific binding antibody. Such a technique is described in Larrick *et al.*, *Biotechnology*, 7:394 (1989).

The monoclonal antibodies and fragments thereof employed as the pTarg component include chimeric antibodies, e.g., humanized versions of murine monoclonal antibodies. Such humanized antibodies may be prepared by known techniques, and offer the advantage of reduced immunogenicity when the antibodies are administered to humans. In one embodiment, the

humanized monoclonal antibody comprises the variable region of a murine antibody (or just the antigen binding site thereof) and a constant region derived from a human antibody. Alternatively, a humanized antibody fragment may comprise the antigen binding site of a murine monoclonal antibody and a variable region fragment (lacking the antigen-binding site) derived from a human antibody. Procedures for the production of chimeric and further engineered monoclonal antibodies include those described in Riechmann *et al.* (*Nature* 332:323, 1988), Liu *et al.* (*PNAS* 84:3439, 1987), Larrick *et al.* (*Bio/Technology* 7:934, 1989), and Winter and Harris (*TIPS* 14:139, May 1993). Procedures to generate antibodies transgenically can be found in GB 2,272,440, US Patent Nos. 5,569,825 and 5,545,806 and related patents claiming priority therefrom, all of which are incorporated by reference herein.

In a further embodiment of the invention, the targeting part (pTarg) of a cytotoxic chimeric polypeptide is a tumor homing peptide. Such a tumor homing peptide include any homing sequence described by Ellerby *et al.*, in example V, VI, VII, VIII of PCT/US00/01602, the entire disclosure of which is relied upon and incorporated by reference herein.

In preferred embodiments of the invention, the chimeric polypeptide has the sequence CNGRCGG-HFRIGCRHSRIG, or CNGRCGG-D[HFRIGCRHSRIG], or CNGRCGG-Vpr52-96, or CNGRCGG-DKRTQFWYFPGN, or CNGRCGG-D[DKRTQFWYFPGN], or ACDCRGDCFCGG-HFRIGCRHSRIG, or ACDCRGDCFCGG-D[HFRIGCRHSRIG], or ACDCRGDCFCGG-Vpr52-96, or ACDCRGDCFCGG-DKRTQFWYFPGN, or ACDCRGDCFCGG-[DKRTQFWYFPGN], or M350/ScFv-HFRIGCRHSRIG, or M350/ScFv-D[HFRIGCRHSRIG] or M350/ScFv-Vpr52-96, or M350/ScFv-DKRTQFWYFPGN, or or M350/ScFv- D[DKRTQFWYFPGN].

Chimeric polypeptides of the invention can be generated by a variety of conventional techniques. Such techniques include those described in B. Merrifield, *Methods Enzymol.* 289:3-13, 1997; H. Ball and P. Mascagni, *Int. J. Pept. Protein Res.* 48:31-47, 1996; F. Molina *et al.*, *Pept. Res.* 9:151-155, 1996; J. Fox, *Mol. Biotechnol.* 3:249-258, 1995; and P. Lepage *et al.*, *Anal. Biochem.* 213: 40-48, 1993.

Peptides can be synthesized on a multi-channel peptide synthesizer using classical Fmoc-based and pseudopeptide synthesis. In one embodiment of the invention, Vpr52-96, Vpr71-96 and Vpr 71-82 and all the Tox, Save and TARG peptides described in Table I, II, III, are

synthesized by solid phase peptide chemistry. After cleavage from the resin, the peptides are purified and analyzed by reverse-phase HPLC. The purity of the peptides is typically above 98% according to HPLC trace. The integrity of each peptide can be controlled by matrix Assisted Laser Desorption Time of Flight spectrometry. To avoid rapid degradation of the peptides in biological fluids, one or several amide bonds could be advantageously replaced by peptide bond isosters like retro-inverso (NH-CO), methylene amino (CH₂-NH), carba (CH₂-CH₂) or carbaza (CH₂-CH₂-N(R)) bonds.

Alternatively, the chimeric polypeptides of the invention can be prepared by subcloning a DNA sequence encoding a desired peptide sequence into an expression vector for the production of the desired peptide. The DNA sequence encoding the peptide is advantageously fused to a sequence encoding a suitable leader or signal peptide. Alternatively, the DNA fragment may be chemically synthesized using conventional techniques. The DNA fragment can also be produced by restriction endonuclease digestion of a clone of, for example HIV-1, DNA using known restriction enzymes (New England Biolabs 1997 Catalog, Stratagene 1997 Catalog, Promega 1997 Catalog) and isolated by conventional means, such as by agarose gel electrophoresis.

In another embodiment, the well known polymerase chain reaction (PCR) procedure can be employed to isolate and amplify a DNA sequence encoding the desired protein or peptide fragment. Oligonucleotides that define the desired termini of the DNA fragment are employed as 5' and 3' primers. The oligonucleotides can contain recognition sites for restriction endonucleases, to facilitate insertion of the amplified DNA fragment into an expression vector. PCR techniques are described in Saiki *et al.*, Science 239:487 (1988); Recombinant DNA Methodology, Wu *et al.*, eds., Academic Press, Inc., San Diego (1989), p. 189-196; and PCR Protocols: A Guide to Methods and Applications, Innis *et al.*, eds, Academic Press., (1990). It is understood of course that many techniques could be used to prepare polypeptide and DNA fragments, and that this embodiment in no way limits the scope of the invention.

Several methods can be used to link TARG to TOX and TARG to SAVE, depending on the particular chemical characteristics of the molecules. For example, methods of linking haptens to carrier proteins as used routinely in the field of applied immunology. In one embodiment, a premade a PTPC regulatory molecule (TOX or SAVE) can be conjugated to an antibody as antibody fragment (pTarg) using, for example, carbodiimide conjugation.

Carbodiimides comprise a group of compounds that have the general formula $R-N^+C=N-R$, where R and R can be aliphatic or aromatic, and are used for synthesis of peptide bonds. The preparative procedure is simple, relatively fast, and is carried out under mild conditions.

Cardodiimide compounds attack carboxylic groups to change them into reactive sites for free amino groups. Carbodiimide conjugation has been used to conjugate a variety of compounds for the production of antibodies.

The water soluble carbodiimide, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC) can be useful for conjugating a PTPC regulatory molecule (TOX or SAVE) to an antibody or antibody fragment molecule. Such conjugation requires the presence of an amino group, which can be provided, for example, by a PTPC regulatory molecule (TOX or SAVE), and a carboxyl group, which can be provided by an antibody or antibody fragment.

In addition to using carbodiimides for the direct formation of peptide bonds, EDC also can be used to prepare active esters, such as N-hydroxysuccinimide (NHS) ester. The NHS ester, which binds only to amino groups, then can be used to induce the formation of an amide bond with the single amino group of the oxorubicin. The use of EDC and NHS in combination is commonly used for conjugation in order to increase yield of conjugate formation.

Other methods for conjugating a PTPC regulatory molecule (TOX or SAVE) to an antibody or antibody fragment also can be used. For example, sodium periodate oxidation followed by reductive alkylation of appropriate reactants can be used, as can glutaraldehyde crosslinking. However, it is recognized that, regardless of which method of producing a chimeric polypeptide of the invention is selected, a determination must be made that an antibody or antibody fragment maintains its targeting ability and that a PTPC regulatory molecule (TOX or SAVE) maintains its activity.

The chimeric polypeptide of the invention may further incorporate a specifically non-cleavable or cleavable linker peptide functionally interposed between the PTPC regulatory molecule (TOX or SAVE) (pTarg) and the antibody or antibody fragment (pTox). Such a linker peptide provides by its inclusion in the chimeric construct, a site within the resulting chimeric polypeptide that may be cleaved in a manner to separate the intact PTPC regulatory molecule (TOX or SAVE) from the intact antibody or antibody fragment. Such a linker peptide may be, for instance, a peptide sensitive to thrombin cleavage, factor X cleavage, or other peptidase

cleavage. Alternatively, where the chimeric polypeptide lacks methionine, the antibody or antibody fragment may be separated by a peptide sensitive to cyanogen bromide treatment. In general, such a linker peptide will describe a site, which is uniquely found within the linker peptide, and is not found at any location in either of the TARG, TOX or SAVE fragment constituting the chimeric polypeptide.

Compositions comprising an effective amount of a chimeric polypeptide of the present invention, in combination with other components, such as a physiologically acceptable diluent, carrier, or excipient, are provided herein. The chimeric polypeptide can be formulated according to known methods used to prepare pharmaceutically useful compositions. They can be combined in admixture, either as the sole active material or with other known materials suitable for a given indication, with pharmaceutically acceptable diluents (e.g., saline, Tris-HCl, acetate, and phosphate buffered solutions), preservatives (e.g., thimerosal, benzyl alcohol, parabens), emulsifiers, solubilizers, adjuvants and/or carriers. Suitable formulations for pharmaceutical compositions include those described in *Remington's Pharmaceutical Sciences*, 16th ed. 1980, Mack Publishing Company, Easton, PA.

In addition, such compositions can be complexed with polyethylene glycol (PEG), metal ions, or incorporated into polymeric compounds such as polyacetic acid, polyglycolic acid, hydrogels, dextran, etc., or incorporated into liposomes, microemulsions, micelles, unilamellar or multilamellar vesicles, erythrocyte ghosts or spheroblasts. Such compositions will influence the physical state, solubility, stability, rate of *in vivo* release, and rate of *in vivo* clearance, and are thus chosen according to the intended application.

The compositions of the invention comprising the chimeric polypeptide can be administered in any suitable manner, e.g., topically, parenterally, or by inhalation. The term "parenteral" includes injection, e.g., by subcutaneous, intravenous, or intramuscular routes, also including localized administration, e.g., at a site of disease or injury. Sustained release from implants is also contemplated. One skilled in the pertinent art will recognize that suitable dosages will vary, depending upon such factors as the nature of the disorder to be treated, the patient's body weight, age, and general condition, and the route of administration. Preliminary doses can be determined according to animal tests, and the scaling of dosages for human administration is performed according to art-accepted practices.

Compositions comprising nucleic acids in physiologically acceptable formulations are also contemplated. DNA may be formulated for injection, for example.

In one of its most general applications, the invention relates to a recombinant vector incorporating a DNA segment having a sequence encoding the chimeric polypeptide of the invention. For the purposes of the invention, the term "chimeric polypeptide" is defined as including any polypeptide where at least a portion of a viral apoptotic peptide is coupled to at least a portion of an antibody or antibody fragment. The coupling can be achieved in a manner that provides for a functional transcribing and translating of the DNA segment and message derived therefrom, respectively.

The vectors of the invention will generally be constructed such that the chimeric polypeptide encoding sequence is positioned adjacent to and under the control of an effective promoter. In certain cases, the promoter will comprise a prokaryotic promoter where the vector is being adapted for expression in a prokaryotic host. In other cases, the promoter will comprise a eukaryotic promoter where the vector is being adapted for expression in a eukaryotic host. In the later cases, the vector will typically further include a polyadenylation signal position 3' of the carboxy-terminal amino acid, and within a transcriptional unit of the encoded chimeric polypeptide. Promoters of particular utility in the vectors of the invention are cytomegalovirus promoters and baculovirus promoters, depending upon the cell used for expression. Regardless of the exact nature of the vector's promoters, the recombinant vectors of the invention will incorporate a DNA segment as defined below.

A recombinant host cell is also claimed herein, which incorporates a vector of the invention. The recombinant host cell may be either a eukaryotic cell or a prokaryotic host cell. Where a eukaryotic cell is used, a Chinese Hamster Ovary (CHO) cell has utility. In another embodiment, when used in combination with a baculovirus promoter, the insect cell lines SF9 or SF21 can be used.

This invention will be described in greater detail in the following Examples.

EXAMPLE 1

Obtaining the murine monoclonal antibody (Ac M350)

Human fetal cells were chosen as a source of immunization. It was the well-known similarities between fetal and tumoral antigens which inspired us to use fetal cells as a source of immunization to produce monoclonal antibodies directed against the epitopes present on tumoral cells. Oncofetal antigens are glycoproteins which are present during intra-uterine life; they disappear at birth and can be re-expressed in pathological situations, particularly in malignant tumors. There are many examples of this antigen community, the best known models being fetoprotein which is associated with 70% of liver tumors, and <<embryo tumor antigens>>, which is often used in human clinical practice and which is a monitoring parameter for patients suffering from cancers of the digestive tract.

A. M350 clone production

These fetal cells were obtained from the sterile removal of the mammary buds of 25-week old female fetuses. Once the buds had been mechanically dissociated into 0.5 mm³ fragments, the cells were resuspended in a Dulbecco medium modified with collagenase and hyaluronidase at 37°C and shaken for between 30 minutes and 4 hours after being monitored under the microscope. As soon as organoids appear, the cells were deposited onto Ficoll, washed, then cultured in a calcium-free DMEM-F12 medium, in hepes, insulin, choleric toxin, cortisol. Once the cells were subcultured once a week. Using this technique the cells duplicated 10 to 20 times giving sufficient cells for immunization purposes.

Balb/c mice were immunized four times, intraperitoneally. The fusion was achieved according to the classical technique of Kohler and Milstein. The screening was done with fetal mammary cells, adult mammary cells and breast tumors. Several clones appeared and one, M350 clone, was particularly tested on breast tumors and normal breast tissues. 150 tumor sections were tested: (i.e.) infiltrating intra-canalar and intra-lobular adenocarcinomas, infiltrating lobular adenocarcinomas. Tests were performed using an immunoenzymatic technique with alkaline phosphatase. All the tumors tested positive whereas the normal tissues taken from mammary

samples tested in parallel were negative for weakly positive. Each slide of normal tissue contained lobular type epithelial structures and cavities inside the paleal tissue.

B. Other Hybridomes

Obtaining new murine monoclonal antibodies against associated breast tumor antigens.

In this technology, C57/B16 mice were immunized four times, intraperitoneally, with a mixture of three different breast tumor cell lines (MCF7, MDA, ZR75-1). After fusion and screening the specificity was studied on normal breast tissues and malignant tumors, other tumor samples and peripheral blood cells. The Monoclonal antibodies showing surface tumor labeling were chosen.

EXAMPLE 2

A Cell lines and viruses

The insect cells derived from ovarian tissue of *Spodoptera frugiperda* (Sf9 insect cells, Vaughn et coll., 1977) and insect cells derived from *Trichoplusia ni* (High Five insect cells) were maintained at 28°C in TC100 medium supplemented with 5% fetal calf serum and were used for the propagation of recombinant baculoviruses and for the production of recombinant proteins. The recombinant baculoviruses are obtained after co-transfection of insect cells with baculovirus viral DNA (Baculogold, Pharmingen) and recombinant transfer vector DNA.

B. Recombinant transfer vector: pVL-PS-gp671

The recombinant transfer vector pVL-PSgp671 derived from transfer vector pVL1392 (Invitrogen) is used as transfer vector to generate recombinant viruses. It includes from 5' to 3' : the peptide signal sequence of gp67 baculovirus glycoprotein, the sequence coding for a His(6)-Tag, the recognition sequence for the Xa Factor, a polylinker region for subcloning the scFv sequence, a link-sequence: GGC required for the covalent association between cytotoxic peptides and ScFv.

The signal peptide sequence from gp67 was added by insertion of a PCR product of gp67 (obtained by PCR from a commercial pcGP67-B plasmid as a template and the PSgp67-Back and PSgp67-For as primers) at the *Bg/II* site of the pVL1392 plasmid. The sequence coding for the His(6)-Tag sequence and the recognition sequence for the Xa factor were then added by using

insertion of oligonucleotides at the 3' end of the gp67 sequence. By the same way the sequence of the peptide motif required for the covalent association between cytotoxic peptides and ScFv: (-Gly-Gly-Cys) was added at the 3' part of the polylinker (the first G is encoded by the last nucleotide of the XmaI site).

Insertion at BamHI and BglII of overlapping primers:

Th1: GAT CCC ATC ATC ACC ACC ACC AC (BamHI-His(6))

Th2: ATT GAA GGA AGA GAATTC CCATG (Factor Xa cleavage -EcoRI-NcoI)

Th3: GCT GCA GCC CGG GGG ATG TTA AA (PstI -XmaI -GGG - STOP- BamHI)

Th4: CTT CCT TCA ATG TGG TGG TGG TGA TGA TGG (link between Th1 Th2)

Th5: GGG CTG CAG CCA TGG GAA TTC T (link between Th2 and Th3)

Th6: GAT CTT TAA CAT CCC CC (link between Th3 and pVL, -pg67)

C Synthesis of ScFv DNA fragment

VH and VL regions of M350:

Total RNA isolated from M350 hybridoma have been used as a template for a reverse transcription using oligo (dT) as primers (Reverse Transcription IBI Fermentas). A PCR realized with those cDNAs and specific primers (mouse Ig-Prime-Kit, Novagen) have led to the selective amplification of VH and VL chains. These regions are then cloned in "blunt" in pST-Blue 1 plasmid and sequenced.

VH and VL regions of other hybridomas:

Total RNA isolated from selected hybridoma was used as a template for a reverse transcription using oligo (dT) (Reverse Transcription IBI Fermentas). A PCR with specific primers (mouse Ig-Prime-Kit, Novagen) led to the selective amplification of VH and VL chains. These products are then cloned in pGEMT (TA cloning System from PROMEGA) vector and sequenced. Three new VH and VL sequences were determined from clone therap.99B3 (**Figure 3**), clone therap.88E10 (**Figure 4**), and therap.152C3 (**Figure 5**).

Obtention of the ScFv-transfer vector:

VH-link-VL chimeric DNA were done by fusion-PCR in two steps (**Figure 12**). The first

step added a link-sequence (Gly-Gly-Gly-Gly-Ser) at the 3' of the VH chain and at the 5' end of the VL chain respectively. The second step was a PCR fusion leading to the chimeric DNA: VH-link-VL. The set of primers used in this second step brings a 5' -EcoRI and a 3'-XmaI sites to VH and VL respectively that will be used for the subcloning of the final product in pVL-PSgp671 vector (**Figure 13**).

D Cotransfection and purification of recombinant baculoviruses

Sf9 cells were cotransfected with viral DNA (BaculoGold ; Pharmingen) and recombinant transfer vector DNA (pVL-PSgp671-ScFv) by the lipofection method (Feloner and Ringold, 1989) (DOTAP; Roche). Screening and purification of recombinant viruses were carried out by the common procedure described by Summers and Smith (Summers and Smith, 1987). The recombinant virus was named BAC-PSgp671-scFv and amplified to constitute a viral stock with an MOI of 10^8 .

E Analysis of recombinant proteins

Infected cells were collected, washed with cold phosphate-buffered saline (PBS) and resuspended in sample reducing buffer (Laemmli, 1970). After boiling (100°C for 5 min), proteins samples were resolved by 12.5% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) under denaturing conditions (Laemmli, 1970). The apparent molecular weight of the protein was checked by coomassie blue staining or the proteins were transferred onto a nitrocellulose filter (Schleicher and Schuell ; BAS 85, 0.45µm) with a semidry blotter apparatus (Ancos). The nitrocellulose membrane was then stained with Ponceau Red (Sigma) and subsequently blocked with a solution of Tris-saline buffer (0.05 M Tris-HCl pH7.4, 0.2 M NaCl) containing 0.05% Tween 20 and 5% non fat milk (TS-sat). ScFv was detected using a mouse monoclonal antibody raised against His(6)-Tag (SIGMA) as primary antibody and a sheep anti-mouse immunoglobulin G (IgG)- horseradish peroxidase conjugate as secondary antibody (1; 3000 Amersham). The immunoreactive bands were visualized by using ECL reagents as described by the manufacturer (Amersham).

F Protein production and purification

To obtain viral stock, Sf9 insect cells cultured in IPL41 medium and 5% FCS are infected in exponential phase with the recombinant baculoviruses at MOI1. After a 7-day incubation period at 28° in IPL41 medium with 5% FCS, the supernatant is harvested by centrifugation at 8000 RPM during 15 min. Then High-five insect cells cultured in Xpress media (Biowhitaker) are infected with recombinant baculovirus in exponential phase at MOI 10, following 1h30 of infection High Five cells were harvested by centrifugation and resuspended in Xpress media without serum. After a 4-day period of incubation at 28°C, the supernatant is harvested by centrifugation at 8000 RPM during 15 min. These supernatants are then concentrated by two rounds of ammonium sulfate precipitation. The precipitate obtained by sedimentation is dialyzed during 12 hours and purified using batch of Ni-NTA agarose beads as described by the manufacturer (Qiagen). After dialysis (2 days, PBS, 4°C) and analysis by Coomassie staining purified proteins were used for the covalent association with cytotoxic peptides.

EXAMPLE 3

Method of coupling ScFv to pTox

The peptide was assembled using Fmoc solid phase peptide synthesis, after the last Fmoc deprotection a propionyloxy succinimide ester was allowed to react, in the presence of diisopropyl ethylamine, with the alpha amino group of the peptide. At the end of the reaction (30 min) the peptide resin was washed with methylene chloride and the peptide was classically cleaved and deprotected under acidic conditions. The activated peptide was then purified by HPLC and its integrity was confirmed by mass spectrometry. The activated peptide was then allowed to react with the ScFv with peptide in a molar ratio of 10:1 (pH7, PBS, glass tube over agitation for 3 hours at room temperature). Then, dialysis was done for 48h against PBS at 4°C. Four Tox peptides were coupled to ScFv using this method:

Tox 11	ScFv-M350-Jac5 (Vpr71-96[C761])
Ctr1Tox11I	ScFv-M350-Jac5M (Vpr71-96[C76S;R73,80A])
Tox 12	ScFv-Vpr52-96[C76S]
Ctr1Tox12	ScFv -Vpr52-96[C76S ; R73A; R80A]

EXAMPLE 4**Examples of Targ-Tox or Targ-Save structures**

All the Tox peptides can have a facultative N-terminal biotin and a facultative C-terminal amide function. Tox0 is a Tox peptide which does not necessarily require an association with a Targ. Tox1, Tox2, Tox 5, Tox6, Save1, Save2 and their respective control can possess a facultative gly-gly- (-GG-) linker between the Targ and the Tox/Save motif.

Tox0	Biot-DTWTGVEALIRILQQLLFHFRIGCRHSRIGIIQQRRTNRNGASKS
CtrlTox0	Biot-DTWTGVEALIRILQQLLFHFAIGCRHSAIGIIQQRRTNRNGASKS

Tox1	Biot- CNGRC-GG-HFRIGCRHSRIG
CtrlTox1	Biot- CNGRC-GG-HFAIGCRHSAIG
CtrlTox1	Biot-CNGRC-GG-CNGRC
CtrlTox1	Biot-GG-HFRIGCRHSRIG
CtrlTox1	Biot-CNGRC-GG-Scramble
CtrlTox1	Biot-KETWWETWWTEW-GG-HFRIGCRHSRIG

Tox2	Biot-ACDCRGDCFC-GG-HFRIGCRHSRIG
CtrlTox2	Biot- ACDCRGDCFC-GG-HFAIGCRHSAIG

Tox5

Tox5	Biot-CNGRC-GG-DKRTQFWRYFPGN (hANT2m)
CtrlTox5	Biot-CNGRC-GG-DKRTQFWRYFAGN (hANT2)
CtrlTox5	Biot-CNGRC-GG-DRHKQFWRYFPGN (hANT1m)
CtrlTox5	Biot-CNGRC-GG-DKHTQFWRYFPGN (hANT3m)
CtrlTox5	Biot-GG-DKRTQFWRYFPGN (hANT2m)
CtrlTox5	Biot-GG-DRHKQFWRYFPGN (hANT1m)
CtrlTox5	Biot-GG-DKHTQFWRYFPGN (hANT3m)
CtrlTox5	Biot-CNGRC-GG-Scramble

Tox6

Tox6	Biot-ACDCRGDCFC-GG-DKRTQFWRYFPGN (hANT2m)
CtrlTox6	Biot-ACDCRGDCFC-GG-DKRTQFWRYFAGN (hANT2)
CtrlTox6	Biot-ACDCRGDCFC-GG-DRHKQFWRYFPGN (hANT1m)
CtrlTox6	Biot-ACDCRGDCFC-GG-DKHTQFWRYFPGN (hANT3m)
CtrlTox6	Biot-ACDCRGDCFC-GG
CtrlTox6	Biot-ACDCRGDCFC-GG-Scramble

Tox 11

Tox 11	ScFv-M350-Jac5(Vpr71-96[C76])
Ctr1Tox11	ScFv-M350-Jac5M(Vpr71-96[C76;R73,80A])

Save1

Save1	Biot-RKKRRQRRR-DKRTQFWRYFAGN (hANT2)
Ctr1Save1	Biot-RKKRRQRRR-DKRTQFWRYFPGN (hANT2m)
Ctr2Save1	Biot-RKKRRQRRR-DRHKQFWRYFAGN (hANT1)
Ctr3Save1	Biot-RKKRRQRRR-DKHTQFWRYFAGN (hANT3)
Ctr4Save1	Biot-RKKRRQRRR
Ctr5Save1	Biot-RKKRRQRRR-Scramble

Save2

Save 2	Biot-RKKRRQRRR-LASGGAAGATSLCFVYPL (hANT[117-134])
Ctr1Save2	Biot-RKKRRQRRR-GAWSNVLRGMGGAFVLVLY (ANTTM6[271-289])
Ctr2Save2	Biot-RKKRRQRRR-scramble

EXAMPLE 5**Evaluation of mitochondrial and nuclear parameters of Apoptosis in cells (cell lines) and cell-free systems****A. Cells**

MCF-7, MDA-MB231, COS and HeLa cells are cultured in complete culture medium (DMEM supplemented with 2 mM glutamine, 10% FCS, 1 mM Pyruvate, 10 mM Hepes and 100 U/ml penicillin/streptomycin). Jurkat cells expressing CD4 and stably transfected with the human Bcl-2 gene or a Neomycin (Neo) resistance vector [Aillet, *et al.*, 1998 J. Virol. 72:9698-9705] only were kindly provided by N. Israel (Pasteur Institute, Paris). Neo and Bcl-2 U937 cells [Zamzami *et al.*, 1995 J. Exp. Med], and CEM-C7 cells are cultured in RPMI 1640 Glutamax medium supplemented with 10% FCS, antibiotics, and 0.8 µg/ml G418.

The cell tests that have been implemented determine the pathway (intracellular penetration, then subcellular localization) of the candidates, and the apoptotic status ($\Delta\psi_m$, activation and relocalization of cell death effectors, content in nuclear DNA) of the target cell. In order to determine these parameters it is necessary to use fluorescent probes to label the cells and/or the candidates molecules and to implement the following two analytical procedures : multi-parameter cytofluorimetry and fluorescent microscopy. As far as neuroprotection is

concerned, tests were carried out on primary cultures of cortical neuronal cells from mice embryos. As far as cardioprotection is concerned, tests were carried out on primary cultures of cardiomyocytes from mice embryos.

- **Intra-cellular pathway tests:** the TARG-TOX ou TARG-SAVE peptides coupled either with biotin (detected using fluorochromes conjugated with streptavidin ; or by ligand-blot after subcellular fractioning) or with FITC (detected by direct observation of living cells, videomicroscopy and image analysis) are added to the cells. It possible to favor the TOX or SAVE mitochondrial routing by inserting mitochondrial addressing signals (the Apoptosis Inducing Factor or ornithin transcarbamylase, for example). Similarly, the mitochondrial routing is evaluated after modifying sequences and certain lateral chains (phosphorylations, methylations), then replacing the peptides by peptidomimetics.

- **Multi-parameter analysis of apoptosis** on tumoral and endothelial cell lines, and primary neurons. Fluorescents probes will be used to mesure the state of the mitochondrial transmembrane potential (JCI, DioC6, mitoTrackers) and nuclear condensation (Hoescht). Similarly, the post-mitochondrial parameters of apoptosis are evaluated using classical hypoploidy tests and cell surface labeling with annexin V-FITC.

In this type of tests, we evaluate either the cytotoxic potential of the TARG-TOX, i.e. their capacity to kill (via a mitochondrial effect) tumoral ou endothelial cell lines (the best TARG-TOX must also kill over-expressing Bel-2 cell lines); or the cytoprotective potential of the TARG-SAVE when the neurons are subjected to different apoptogenic treatments.

B. Apoptosis Modulation

PBS-washed cells ($1-5 \times 10^5$ /ml) are incubated with (1 to 5 μ M) of pTarg-pTox in complete culture medium supplemented or not with cyclosporin A (CsA; 1 μ M), bongkreikic acid (BA; 50 μ M), and/or the caspase inhibitors N-benzyloxycarbonyl-Val-Ala-Asp.fluoromethylketone (Z-VAD.fmk; 50 μ M; Bachem Bioscience, Inc.), Boc-Asp.fluoromethylketone (Boc-D.fmk), or N-benzyloxycarbonyl-Phe-Ala-fluoromethylketone (Z-FA.fmk; all used at 100 μ M added each 24 h; Enzyme Systems). During exposure to pTarg-pTox, human primary PBLs from healthy donors, purified with Lymphoprep (Pharmacia), are cultured in RPMI 1640 Glutamax medium without any addition of serum. In contrast, PHA blasts (24 h of 1 μ g/ml PHA-P [Wellcome Industries]; 48 h with 100 U/ml human recombinant IL-2 [Boehringer Mannheim]) are cultured with 10% FCS.

C. Cytofluorimetric Determinations of Apoptosis-associated Alterations in Intact Cells

For cytofluorometry, the following fluorochromes are employed: 3,3'-dihexyloxacarbocyanine iodide (DiOC(6)3; 40 nM) for mitochondrial transmembrane potential ($\Delta\Psi$ m) quantification, hydroethidine (4 μ M) for the determination of superoxide anion generation, and propidium iodide (PI; 5 μ M) for the determination of viability (Zamzami, N., *et al.*, 1995. J. Exp. Med. 182:367-377). The frequency of subdiploid cells is determined by PI (50 μ g/ml) staining of ethanol-permeabilized cells treated with 500 μ g/ml RNase (Sigma Chemical Co.; 30 min, room temperature [RT]) in PBS, pH 7.4, supplemented with 5 mM glucose (Nicoletti, I. *et al.*, 1991. J. Immunol. Methods. 139:271-280).

D. Fluorescence staining of life cells and immunofluorescence

For the assessment of mitochondrial and nuclear features of apoptosis, cells cultured on a cover slip are incubated with the $\Delta\Psi$ m-sensitive dyes chloromethyl-X-rosamine (CMXRos; 50 nM; Molecular Probes, Inc.) or 5,5',6,6'-tetrachloro-1,1', 3,3'-tetraethylbenzimidazolylcarbocyanine iodide (JC-1, 2 μ M, Molecular Probes), the $\Delta\Psi$ m-insensitive dye Mitotracker green (1 μ M; Molecular Probes, Inc.), and/or Hoechst 33342 (2 μ M, Sigma) for 30 min at 37°C in complete culture medium (Marzo, *et al.* 1998. Science. 281:2027-2031).

E. For in situ determinations of pTarg-pTox internalisation

For *in situ* determinations of TARG-(MLS)-TOX/SAVE internalisation, cells are incubated at different times with TARG-(MLS)-TOX/SAVE, and then cells are fixed with 4% paraformaldehyde and 0.19% picric acid in PBS (pH 7.4) for 1 h at RT. Fixed cells are permeabilized with 0.1% SDS in PBS at RT (for 5 min), blocked with 10% FCS, and stained with an mAb specific for hexa-histidine tag (clone HIS-1, IgG2a, SIGMA) revealed by a goat anti-mouse PE conjugate [Southern Biotechnology Associates, Inc.], Hsp60 (mAb H4149 [Sigma Chemical Co.], revealed by a goat anti-mouse IgG1 FITC conjugate), cytochrome c oxidase (COX; mAb 20E8-C12 [Molecular Probes, Inc.], revealed by a goat anti-mouse IgG2a FITC conjugate), or when the Targ is a biotinylated peptide, a streptavidin-PE reagent is added 30 min. followed by detection of the fluorescence intensity by fluorescence (and/or confocal) microscopy.

F. Assessment of mitochondrial parameters in vitro

Mitochondria are purified from rat liver, as described (Costantini *et al.*, 1996), and resuspended in 250 mM sucrose + 0.1 mM EGTA + 10 mM -tris[hydroxymethyl]methyl-2-Aminoethanesulfonic acid, pH=7.4). For the induction of PT, mitochondria (0.5 mg protein per ml) are resuspended in PT buffer (200 mM sucrose, 10 mM Tris-MOPS (pH 7.4), 5 mM Tris-succinate, 1 mM Tris-phosphate, 2 μ M rotenone, and 10 μ M EGTA-Tris), and monitored in an F4500 fluorescence spectrometer (Hitachi, Tokyo, Japan) for the 90° light scattering of light (545 nm) to determine large amplitude swelling after addition of 2 mM atractyloside (Atr), 1 μ M cyclosporin A (CsA; Novartis, Basel, Switzerland), 5 μ M CaCl_2 , and/or 0.5 to 20 μ M of pTarg-pTox or pTarg-pSave. For the determination of the $\Delta\Psi_m$, mitochondria (0.5 mg protein per ml) are incubated in a buffer supplemented with 1 μ M rhodamine 123 (Molecular Probes, Eugene, OR) and the dequenching of rhodamine fluorescence (excitation 505 nm, emission 525 nm) is measured as described (Shimizu *et al.*, 1998). Supernatants from mitochondria (6800 g for 15min; then 20 000 g for 1 h; 4°C) are frozen at -80°C until determination of apoptogenic activity on isolated nuclei, DEVD-afc cleaving activity, and immunodetection of cytochrome c and AIF. Cytochrome c and AIF are detected by means of a monoclonal antibody (clone 7H8.2C12, Pharmingen) and a polyclonal rabbit anti-serum (Susin *et al.* 1999) respectively.

Swelling of isolated mitochondria**Table F1 :**

Tox0, Tox1, Tox5, Tox6 induce permeability transition pore (PTP) opening

Name of molecules 5 μ M	Induction of Mitochondrial swelling (sw) +++ rapid sw ; ++ low sw ; + very low sw ; - no sw t 20 min
-	-
Tox0	+++
Tox1	++
CtrlTox1	-
Ctrl2Tox1	-
Ctrl3Tox1	+
Ctrl4Tox1	-
Tox5	++
Tox6	++

Table F2:

Save 1 and Save2 inhibit atractyloside-induced PTP opening

Name of molecules	Mitochondrial swelling (sw) %
-	2
Ca 2+ 100 μ M	100
Atractyloside 600 μ M	110
Save I 5 μ M	2
Save I 5 μ M + Atr 600 μ M	5
Save I 20 μ M	12
Save I 20 μ M + Atr 600 μ M	12
Save II 10 μ M	2
Save II 20 μ M	16
Save II 10 μ M + Atr 600 μ M	16
Save II 20 μ M + Atr 600 μ M	16

G. ANT purification and reconstitution in liposomes

ANT was purified from rat heart mitochondria as previously described (8). After mechanical shearing, mitochondria were suspended in 220 mM mannitol, 70 mM sucrose, 10 mM Hepes, 200 μ M EDTA, 100 mM DTT, 0.5 mg/ml subtilisin, pH7.4, kept 8 min on ice and sedimented twice by differential centrifugations (5 min, 500 x g, and 10 min, 10,000 x g). Mitochondrial proteins were solubilized by 6% [v:v] Triton X-100 (Boehringer Mannheim) in 40 mM K_2HPO_4 , 40 mM KCl, 2 mM EDTA, pH 6.0, for 6 min at RT and solubilized proteins were recovered by ultracentrifugation (30 min, 24,000 x g, 4°C). Then, 2 ml of this Triton X-100 extract was applied to a column filled with 1 g of hydroxyapatite (BioGel HTP, BioRad), eluted with previous buffer and diluted [v:v] with 20 mM MES, 200 μ M EDTA, 0.5% Triton X-100, pH6.0. Subsequently, the sample was separated with a Hitrap SP column using a FPLC system (Pharmacia) and a linear NaCl gradient (0-1M). Proteins concentration was determined using microBCA-assay (Pierce, Rockfall, Illinois). Purified ANT and/or recombinant Bcl-2 were reconstituted in PC/cardioliipin liposomes. Briefly, to prepare liposomes, 45 mg PC and 1 mg cardioliipin were mixed in 1 ml chloroform, and the solvent was evaporated under nitrogen. Dry lipids were resuspended in 1 ml liposome buffer (125 mM sucrose + 10 mM -2-hydroxyethylpiperazine-*N'*-2 ethanesulfonic acid; Hepes, pH 7.4) containing 0.3% n-octyl- β -D-pyranoside and mixed by continuous vortexing for 40 min at RT. ANT (0.1 mg/ml) or recombinant Bcl-2 (0.1 mg/ml) were then mixed with liposomes [v:v] and incubated for 20 min at RT. Proteoliposomes were finally dialysed overnight at 4°C.

H. Pore opening assay

ANT-proteoliposomes are sonicated in the presence of 1 mM 4-MUP and 10 mM KCl (50W, 22sec, Branson sonifier 250) on ice as previously described (28). Then, liposomes were separated on Sepadex G-25 columns (PD-10, Pharmacia) from unencapsulated products. 25 μ l- aliquots of liposomes were diluted to 3 ml in 10 mM Hepes, 125 mM saccharose, pH 7.4, mixed with various concentrations of the proapoptotic inducers and incubated for 1 h at RT. Potential inhibitors of mitochondrial membranes permeabilization such as BA, ATP and ADP, were added to the liposomes 30 min before treatment. After addition of 10 μ l-alkaline phosphatase (5 U/ml, Boehringer Mannheim) diluted in liposomes buffer + 0.5 mM $MgCl_2$, samples were incubated

for 15 min at 37°C under agitation and the enzymatic conversion of 4-MUP in 4-MU was stopped by addition of 150 µl Stop buffer (10 mM Hepes-NaOH, 200 mM EDTA, pH 10). The 4-MU-dependent fluorescence (360/450 nm) was subsequently quantitated (28) using a Perkin Elmer spectrofluorimeter. Atractyloside, a pro-apoptotic permeability transition inducer, was used in each experiment as a standard to determine the 100% response. The percentage of 4-MUP release induced by Vpr-derived peptides or pTarg-ptox was calculated as following :

$$\frac{[(\text{fluorescence of liposomes treated by pTar-pTox} - \text{fluorescence of untreated liposomes}) / (\text{fluorescence of liposomes treated by atractyloside} - \text{fluorescence of untreated liposomes})] \times 100.}$$

ANT pore opening assay:

Table H1 : examples of functional interaction between ANT and Tox or Save constructs.

Tox0 and Tox6 induce ANT-protéoliposomes permeabilisation. Save1 and Save2 block Atractyloside (Atra) -induced ANT-protéoliposomes permeabilisation

molecules	Permeabilisation of ANT - proteoliposomes +++ high UMP release ; ++ UMP release ; + low UMP release ; - no UMP release
-	-
Atra 50µM	+
Atra 100µM	++
Atra 200µM	+++
Tox0 (Biotin-Vpr52-96) 2µM	+++
Tox6 5µM	++
Biotin-Vpr71-96[C76S] 5µM	++
Save1 5µM	-
Atra 200µM + Save1 5µM	-
Save2 5µM	-
Atra 200µM + Save2 5µM	-

I. Binding assays and western blot

Mouse liver mitochondria were isolated as described (zamzami *et al.*, 2000). For the determination of cytochrome C release, supernatants from pTarg-pTox treated mitochondria (6800 g for 15min; then 20 000 g for 1 h; 4°C) were frozen at -80°C until immunodetection of cytochrome c (mouse monoclonal antibody clone 7H8.2Cl2, Pharmingen). For binding assays, purified mitochondria were incubated (250 µg of protein in 100 µl swelling buffer) for 30 min at

RT 5 μ M (binding assay) of pTarg-pTox or biotin-pTarg-pTox. Mitochondria were lysed either after incubation with biotinylated Vpr52-96 (upper panel) or lysed before (lower panel) with 150 μ l of a buffer containing 20 mM Tris/HCl, pH 7.6; 400 mM NaCl, 50 mM KCl, 1mM EDTA, 0.2 mM PMSF, aprotinin (100U/ml), 1% Triton X-100 and 20% glycerol. Such extracts were diluted with 2 volumes of PBS plus 1mM EDTA before the addition of 150 μ l avidin-agarose (ImmunoPure, from Pierce) to capture the biotin-labeled Vpr52-96 complexed with its mitochondrial ligand(s) (2 hours at 4°C in a roller drum). The avidin-agarose was washed batchwise with PBS (5 x 5 ml; 1000 g, 5 min, 4°C), resuspended in 100 μ l of 2 fold concentrated Laemmli buffer containing 4% SDS and 5 mM β -mercaptoethanol, incubated 10 min at RT and centrifuged (1000 g, 10 min, 4°C). Finally, the supernatants were heated at 95°C for 5 min and analysed by SDS-PAGE (12%), followed by Western blot and immunodetection with a rabbit polyclonal anti-serum against human ANT (kindly provided by Dr. Heide H. Schmid; The Hormel Institute, University of Minnesota, MI; Ref).

J. Flow cytometric analysis of purified mitochondria

Mouse liver mitochondria are isolated as described (zamzami *et al.*, 2000). Purified mitochondria are resuspended in PT buffer (200 mM sucrose, 10 mM Tris-MOPS (pH 7.4), 5 mM Tris-succinate, 1 mM Tris-phosphate, 2 μ M rotenone, and 10 μ M EGTA). Cytofluorometric (FACSVantage, Beckton Dickinson) detection is restricted to mitochondria by gating on the FSC/SSC parameters and on the main peak of the FSC-W parameter. Confirmation *a posteriori* of the validity of these double gating is obtained by labeling of mitochondria with the $\Delta\Psi_m$ -insensitive mitochondrial dye MitoTracker[®] Green (75 nM; Molecular Probes; green fluorescence). To determine the percentage of mitochondria having a low $\Delta\Psi_m$, the $\Delta\Psi_m$ -sensitive fluorochrome JC-1 (200 nM; 570-595 nm) is added 10 min before CCCP or pTarg-pTox molecules. Percentage of mitochondria having a low $\Delta\Psi_m$, is determined in dot-plot FSC/FL-2 (red fluorescence) windows.

K. Cell-free system of apoptosis

AIF activity in the supernatant of mitochondria is tested on HeLa cell nuclei, as described (Susin *et al.*, 1997b). Briefly, AIF-containing supernatants of mitochondria are added to purified HeLa nuclei (90 min, 37°C), which are stained with propidium iodide (PI; 10 μ g/ml; Sigma Chemical Co.) and analyzed in an Elite II cytofluorometer (Coulter) to determine the frequency

of hypoploid nuclei. In some experiments isolated mitochondria, cytosols from Jurkat or CEM cells (prepared as described (Susin *et al.*, 1997a)), and/or pTarg-pTox are added to the nuclei. Caspase activity in the mitochondrial supernatant was measured using Ac-DEVD-amido-4-trifluoromethylcoumarin (Bachem Bioscience, Inc.) as fluorogenic substrate.

L. Purification and reconstitution of PTPC in liposomes

PTPC from Wistar rat brains are purified and reconstituted in liposomes following published protocols (Brenner *et al.*, 1998; Marzo *et al.*, 1998b). Briefly, homogenized brains are subjected to the extraction of triton-soluble proteins, adsorption of proteins to a DE52 resin anion exchange column, elution on a KCl gradient, and incorporation of fractions with maximum hexokinase activity into phosphatidylcholine/cholesterol (5: 1, w:w) vesicles by overnight dialysis. Recombinant human Bcl-2 (1-218) lacking the hydrophobic transmembrane domain (Δ 219-239), produced and purified as described (Schendel *et al.*, 1997) are added during the dialysis step at a dose corresponding to 5% of the total PTPC proteins (approximately 10 ng Bcl-2 per mg lipids). Liposomes recovered from dialysis are ultrasonicated. (120 W) during 7 sec in 5 mM malate and 10 mM KCl, charged on a Sephadex G50 columns (Pharmacia), and eluted with 125 mM sucrose + 10 mM HEPES (pH 7.4). Aliquots (approx. 10^7) of liposomes are incubated during 60 min at RT in 125 mM sucrose + 10 mM HEPES (pH 7.4) in the presence or absence of pTarg-pTox, [52-96]Vpr or atractyloside. Then, liposomes are equilibrated with 3,3'-dihexylocarbocyanine iodide (DiOC₆(3), 80 nM, 20-30 min at RT; Molecular Probes), and analyzed in a FACS-Vantage cytofluorometer (Becton Dickinson, San José, CA, USA) for DiOC₆(3) retention, as described (Brenner *et al.*, 1998; Marzo *et al.*, 1998b).

Triplicates of 5×10^4 liposomes are analyzed and results are expressed as % of reduction of DiOC₆(3) fluorescence, considering the reduction obtained with 0.25% SDS (15 min, RT) in PTPC liposomes as 100% value.

Examples of specific peptides and constructs relating to this invention that can be utilized in carrying out the foregoing techniques are shown in Tables I, II, and III, as well as any chimeric molecule that is a combination between TARG and TOX or TARG and SAVE peptides or peptidomimetics.

EXAMPLE 6

Surface plasmon resonance indicates that Tox0, Tox1, Tox5, Tox6, Save1 binds purified ANT but not purified VDAC.

Methodology.

Sensor Chips SA (streptavidin coated sensor chips) were used for immobilisation of the different peptides. Tox1 was immobilised at a density of 0.7 ng/mm^2 , Tox0 at a density of 3.7 ng/mm^2 , CtrlTox0 at a density of 1.4 ng/mm^2 , Tox5 at a density of 1 ng/mm^2 , Tox6 at a density of 1 ng/mm^2 , Save1 at a density of 1.3 ng/mm^2 , and the control peptide at a density of 0.8 ng/mm^2 . Association and dissociation kinetics of ANT and VDAC interactions were followed at a rate of $10 \text{ }\mu\text{L/min}$ for 10 minutes (5 minutes association and 5 minutes dissociation). The ligand was regenerated with a 1 minute flux of KSCN 3M. The obtained sensorgrams were analysed by the BIAeval 3.1 software using the method of double references (Myszka D.G. 2000. Kinetic, equilibrium and thermodynamic analysis of macromolecular interactions with BIACORE. *Methods Enzymol.* 323:325-340). From the sensorgrams with the ligands were first subtracted the sensorgrams obtained with the corresponding analyte solvents. A second subtraction was performed with the sensorgrams obtained with the control peptide ligand. The control peptide for the Tox and Save peptides was biot-H19C corresponding to the sequence of the β 2-adrenergic receptor (Lebesgue D., Wallukat G., Mijares A., Granier C., Argibay J., and Hoebeke J. (1998) An agonist-like monoclonal antibody to the human β 2-adrenergic receptor. *Eur.J. Pharmacol.* 348:123-133). The control peptide for Tox0 was CtrlTox0.

Results.

Figure 6 shows the interaction between ANT and Vpr for 4 ANT concentrations (6.25 to 50 nM). The sensorgrams were best analysed using the simple Lagmuir model with drifting baseline and resulted in a K_d of 0.15 nM with a R_{max} of 160 ($\chi^2 = 7.24$). The same analysis was performed for the sensorgrams showing the interaction between ANT and Tox1 (**Figure 7**). Studying the VDAC interaction both with Tox0 and Tox1 at VDAC concentrations which were ten times higher (**Figure 8 and 9**), the sensorgrams showed only extremely low association with

the peptide ligand and the obtained curves could not be analysed by the different Langmuir bindings models.

Three other peptides were tested for their interaction with ANT at a concentration of 50 nM (**Figure 10**). Purified ANT recognised Tox5, Tox6, and Save1 with relative affinities of respectively 0.1, 0.7, and 0.01 nM. These values being obtained at only one ANT concentration only give the relative affinity of ANT for the three peptides. Again, the use of 50 nM VDAC to interact with the same peptides did not result in any specific binding as shown in **Figure 11**.

The following references have been cited herein. The entire disclosure of each reference cited herein is relied upon and incorporated by reference herein.

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What is claimed is:

1. Method for inducing or preventing the apoptosis of eukaryotic cells comprising the homing on specific tissue cell population of a chimeric bifunctional molecule able to modulate the activity of permeability transition pore complex (PTPC).
2. A method according to claim 1, wherein said chimeric molecules modulate the activity of the permeability transition pore complex (PTPC) of a specific eukaryotic cell by the regulation of opening or the closing of said pore.
3. A method according to claim 1 or 2, wherein said chimeric molecules comprising at least a first functional molecule and a second functional molecule, wherein said first functional molecule has the function to target specifically a tissue cell population, and the second functional molecule has the function to regulate the apoptosis activity linked to the permeability transition pore complex (PTPC) of said specific cells.
4. A method according to claim 3, wherein said chimeric molecules comprising at least a first functional molecule and a second functional molecule, wherein said first functional molecule has the function to target and to enter specifically in a tissue cell population and the second functional molecule has the function to regulate the apoptosis activity linked to the permeability transition pore complex (PTPC) of said specific cells.
5. A method according to claim 3, wherein said chimeric molecules comprising at least a first functional molecule and a second functional molecule, wherein said first functional molecule has the function to target and to enter specifically in a tissue cell population of interest and the second functional molecule has the function to target specifically and inducing or preventing the death of said cells by apoptosis by the regulation of the

opening or the closing of the permeability transition pore complex (PTPC) of mitochondria or a fragment thereof.

6. A method according to claim 4, wherein said chimeric molecule has the formula:

Targ-Tox,

wherein Tox is a viral or a retroviral apoptotic peptide or a peptidomimetic or a fragment of a protein that interacts with permeability transition pore complex (PTPC) of a specific eukaryotic cell to cause apoptosis of the cell; and Targ is chosen from:

an antibody,

an antibody fragment,

arecombinant antibody fragment,

M350/ScFv,

V461/ScFv,

a homing peptide, and

any peptide chosen in Table III,

wherein said molecule binds and enters the cell specifically.

7. A method according to claim 5, wherein said chimeric molecule has the formula

Targ-Save,

wherein Save is a viral or a retroviral or a cellular antiapoptotic peptide or peptidomimetic or a fragment of protein that interacts with permeability transition pore complex (PTPC) of a specific eukaryotic cell to prevent the apoptosis of the cell with the proviso that when Save peptide is a viral peptide, Save is not vMIA protein of Cytomegalovirus; and

Targ is chosen from:

an antibody,

an antibody fragment,

a recombinant antibody fragment,

M350/ScFv,

a homing peptide, and

any peptide chosen in Table III,

wherein said molecule binds and enters the cell specifically.

8. A method according to anyone of claims 1 to 7, wherein said chimeric molecules comprises a Mitochondrial Localisation Sequence (MLS), which has the function to address specifically the second functional molecule to mitochondrial or intermembrane space-of the mitochondria.
9. A method according to claims 1, 2, 3, 4, 5, 6 and 8, wherein Tox is chosen from the group of peptides of Table I.
10. A method according to claims 1, 2, 3, 4, 5, and 7, wherein Save is chosen from the group of peptides of Table II.
11. A method according to any one of claims 1 to 10, wherein the second functional molecule of said chimeric molecules has the function to interact specifically with ANT of the PTPC of mitochondria also refers to as adenine nucleotide translocator isoforms 1, 2, or 3.
12. A chimeric bifunctional molecule capable to enter specifically in a tissue cell population for induce or prevent death of said cell by apoptosis and comprising at least a first functional molecule covalently linked to a second functional molecule, wherein said first

functional molecule has the function to target and to enter specifically in a tissue cell population of interest and the second functional molecule has the function to target specifically and inducing or preventing the death of said cells by apoptosis by the regulation of the opening or the closing of the permeability transition pore complex (PTPC) of mitochondria or a fragment thereof.

13. A chimeric molecule according to claim 12 which has the formula:

Targ-Tox,

wherein Tox is a viral or a retroviral apoptotic peptide or peptidomimetic or a fragment of a protein that interacts with PermeabilityTransition Pore Complex (PTPC) of a specific eukaryotic cell to cause apoptosis of the cell; and

Targ is chosen from:

an antibody,

an antibody fragment,

a recombinant antibody fragment,

M350/ScFv,

V461/ScFv,

a homing peptide, and

any peptide of Table III,

wherein said molecule binds and enters the cell specifically.

14. A chimeric molecule according to claim 12 which has of the formula

Targ-Save

Wherein Save is a viral or a retroviral or a cellular antiapoptotic peptide or peptidomimetic or a fragment of protein that interacts with Permeability Transition. Pore Complex (PTPC) of a specific eukaryotic cell to prevent apoptosis of the cell, with the proviso that when Save peptide is a viral peptide, Save is not vMIA protein of Cytomegalovirus;

and Targ is chosen from:

an antibody,

an antibody fragment,

a recombinant antibody fragment,

M350/ScFv,

a homing peptide, and

any peptide of Table III,

wherein said molecule binds and enters the cell specifically.

15. A chimeric molecule according to any of claims 12 to 14 comprising a mitochondrial localisation sequence (MLS) which has the function to address specifically the second functional molecule to mitochondrial membranes or intermembrane space.
16. A chimeric molecule according to claims 13 or 15, wherein Tox is chosen from the group of peptides of Table I.
17. A chimeric molecule according to claims 14 and 15, wherein wherein Save is chosen from the group of peptides of Table II.
18. A chimeric molecule according to claims 13, 15 and 16, wherein the Targ and Tox peptides are covalently bonded through a peptide linker comprising 3 to 18 amino acids.

19. A chimeric molecule according to claims 14, 15 and 17, wherein the Targ and Save peptides are covalently bonded through a peptide linker comprising 3 to 18 amino acids.
20. A vector encoding a chimeric molecule as claimed in any one of claims 12 to 19.
21. A hybridoma secreting Targ according to claim 13 or 14 and deposited at the National Collection of Culture and Microorganism (C.N.C.M.) on January 24, 2001, under the accession number n° I 2617.
22. A purified monoclonal antibody encoded by the hybridoma of claim 21.
23. A recombinant host cell comprising a vector as claimed in claim 20.
24. A cancer cell having a tumor associated antigen on the surface thereof to which is bound the chimeric molecule as claimed in any one of claims 12 to 19.
25. A method of determining the presence of a cancer cell having a tumor-associated antigen on the surface thereof in a biological sample comprising :
 - a) contacting a biological sample of interest with a chimeric peptide molecule according to claims 12 to 19 under conditions to permit the binding between the chimeric peptide according to the invention and the antigen on the surface of the cancer cell,
 - b) detecting the binding by usual technique; and
 - c) optionally quantifying the binding detected in step b).
26. A method for inducing death by apoptosis in a tumoral or viral infected cell having a tumor-associated antigen on surface thereof in a biological sample comprising:
contacting a biological sample of interest with a chimeric peptide molecule according to claims 16 or 17 under conditions to permit the binding between the chimeric peptide

according to the invention and the antigen on the surface of the cancer cell and for a time sufficient to allow the entry inside the cell and death cell by apoptosis or viral infected cells.

27. A method for prevent cell death by mitochondrial apoptosis comprising contacting a biological sample of interest with a chimeric molecule, **molecule** according to claims 17 or 19 under conditions to permit the binding between the chimeric molecule according to the invention and the cell of interest and for a time sufficient to allow the entry inside cell of interest and prevent the cell death by apoptosis.
28. A method for prevent cell death according to claim 27, wherein the cells of interest are choosen among the following cell populations: neurons, cardiocytes, and hepatocytes.
29. A method for identifying an active agent of interest that interacts with the activity of the permeability transition pore complex (PTPC) comprising
- a) contacting a biological sample containing cells with permeability transition pore complex (PTPC) with a chimeric peptide according to claims 12 to 19 in the presence of a candidate agent; and
 - b) comparing the binding of the chimeric peptide with the permeability transition pore complex (PTPC) in absence of said agent.
 - c) optionally, testing the activity of said selected agent on a preparation of a cellular extract comprising subcellular elements with the permeability transition pore complex (PTPC).

30. A method for identifying an active agent of interest that interacts with ANT peptide of permeability transition pore complex (PTPC) comprising:
- d) contacting a biological sample containing cells with ANT peptide of permeability transition pore complex (PTPC) with a chimeric peptide according to claims 12 to 19 in the presence of a candidate agent; and
 - e) comparing the binding of the chimeric peptide with the ANT peptide of the permeability transition pore complex (PTPC) in absence of said agent.
 - f) optionally, testing the activity of said selected agent on a preparation of a cellular extract comprising subcellular elements with the ANT peptide of the permeability transition pore complex (PTPC).
31. A method of identification of mitochondrial antigen, said antigen having the capacity to interact with a macromolecule or a molecule or a peptide carrying the characteristic of Tox according to claims 13 or 16.
32. A method of identification of mitochondrial antigen, said antigen having the capacity to interact with a macromolecule or a molecule or a peptide carrying the characteristic of save according to claims 14 or 17.
33. A method of treatment or of prevention of a pathological infection or disease comprising the administration to a patient of the pharmaceutical composition containing at least a chimeric molecule according to any of claims 12 to 19.
34. A pharmaceutical composition comprising at least a chimeric molecule according to claims any of 12 to 19.

Fig. 1A

DNA sequence 10517 b.p. AAGCTTCTACTCG ... AGGCTCTGCG linear

1/21

pACgp67-ScFv461 -> 1-phase Translation

Fig 1B

16/01/1 22:01:48 Page 2

2431/811
AAT TAA ATA GCT TGC GAC GCA ACG TGC ACG ATC TGT GCA CGC GTT CCG GCA CGA GCT TTG ATT GEA ATA AGT TTT TAC GAA CCG ATG ACA
asn och ile ala cys asp ala thr cys thr ile cys ala arg val pro ala arg ala leu ile val ile ser phe tyr glu ala met thr
2521/841
TGA CCC CCG TAG TGA CAA CGA TCA CGC CCA AAA GAA CCG CCG ACT ACA AAA TCA CCG AGT ATG TCG GTG ACG TTA AAA CTA TTA AGC CAC
OPA pro pro pro AMS OFA gln arg ser arg pro lys glu leu pro thr thr lys leu pro ser met ser val thr leu lys leu leu ser his
2611/871
CCA ATC GAC CGT TAG TCG AAT CAG GAC CCG TCG TCG GAG AAG CCG CGA AGT ATG GCG AAT GCA TCG TAT AAC GTG TCG AGT CCG CTC ATT
pro ile asp arg AMS ser asn gln asp arg trp cys glu lys pro arg ser met ala asn ala ser tyr asn val trp ser pro leu ile
2701/901
AGA GCG TCA TGT TTA GAC AAG AAA GCT ACA TAT TTA ATT GAT CCC GAT GAT TTT ATT GAT AAA TTG ACC CTA ACT CCA TAC ACG GTA TTC
arg ala ser cys leu asp lys lys ala thr tyr leu ile asp pro asp asp phe ile asp lys leu thr leu thr pro tyr thr val phe
2791/931
TAC AAT GGC GGG GTT TTG GTC AAA ATT TCC GGA CTG CGA TTG TAC ATG CTG TTA ACG GCT CCG CCC ACT ATT AAT GAA ACT AAA AAT TCC
tyr asn gly gly val leu val lys ile ser gly leu arg leu tyr met leu leu thr ala pro pro thr ile asn glu ile lys asn ser
2881/961
AAT TTT AAA AAA CCG AGC AAG AGA AAC ATT TGT ATG AAA GAA TGC GEA GAA GGA AAG AAX AAT GTC GTC GAC ATG CTG AAC AAC AAG ATT
asn phe lys lys arg ser lys arg asn ile cys met lys glu cys val glu gly lys lys asn val val asp met leu asn asn lys ile
2971/991
AAT ATG CCT CCG TGT ATA AAA AAA ATA TTG AAC GAT TTG AAA GAA AAC AAT GTA CCG CCG GGC GGT ATG TAC ACG AAG AGG TTT ATA CTA
asn met pro pro cys ile lys lys ile leu asn asp leu lys glu asn asn val pro arg gly gly met tyr arg lys arg phe ile leu
3061/1021
AAC TGT TAC ATT GCA AAC GTG GTT TCG TGT GCC AAG TGT GAA AAC CGA TGT TTA ATC AAG GCT CTG ACG CAT TTC TAC AAC CAC GAC TCC
asn cys tyr ile ala asn val val ser cys ala lys cys glu asn arg cys leu ile lys ala leu thr his phe tyr asn his asp ser
3151/1051
AAG TGT GTG GGT GAA GTC ATG CAT CTT TTA ATC AAA TCC CAA GAT GTC TAT AAA CCA CCA AAC TGC CAA AAA ATG AAA ACT GTC GAC AAG
lys cys val gly gly leu met his leu leu ile lys ser gln asp val tyr lys pro pro asn cys gln lys met lys thr val asp lys
3241/1081
CTC TGT CCG TTT GCT GGC AAT TCC AAG GGT CTC AAT CCT ATC TGT AAT TGA ATA ATA AAA CAA TTA TAA ATG CTA AAT TTG TTT TTT
leu cys pro phe ala gly asn cys lys gly leu asn pro ile cys asn tyr OPA ile ile lys gln leu och met leu asn leu phe phe
3331/1111
ATT AAC GAT ACA AAC CAA ACG CAA CAA GAA CAT TTG TAG TAC TAT CTA TAA TTG AAA ACG CGT AGT TAT AAT CGC TGA GGT AAT ATT TAA
ile asn asp thr asn gln thr gln gln glu his leu AMS tyr tyr leu och leu lys thr arg ser tyr asn arg OPA gly asn ile och
3421/1141
AAT CAT TTT CAA ATG ATT CAC AGT TAA TTT CCG ACA ATA TAA TTT TAT TTT CAC ATA AAC TAG ACG CCT TGT CGT CTT CTT CTT CTT ATT
asn his phe gln met ile his ser och phe ala thr ile och phe tyr phe his ile asn AMS thr pro cys arg leu leu leu arg ile
3511/1171
CCT TCT CTT TTT CAT TTT TCT CCT CAT AAA AAT TAA CAT AGT TAT TAT CTT ATC CAT ATA TGT ATC TAC CGT ATA GAG TAA ATT TTT TGT
pro ser leu phe his phe ser pro his lys asn och his ser tyr tyr arg ile his ile cys ile tyr arg ile glu och ile phe cys
3601/1201
TGT CAT AAA CAT ATA TGT CTT TTT TAA TGG GGT GTA TAG TAC CCG TGC GCA TAG TTT TTC TGT AAT TTA CAA CAG TGC TAT TTT CTG GTA
cys his lys tyr ile cys leu phe och trp gly val AMS tyr arg cys ala AMS phe phe cys asn leu gln gln cys tyr phe leu val
3691/1231
GTT CTT CCG AGT GTG CTT TAA TTA TTA AAT TTA TAT AAG CAA TGA ATG TGG GAT CGT CCG TTT TGT ACA ATA TGT TCC CCG CAT AGT
val leu arg ser val leu och leu leu asn leu tyr asn gln OPA ile trp asp arg arg phe cys thr ile cys cys arg his ser
3781/1261
ACG CAG CTT CTT CTA GTT CAA TTA CAC CAT TTT TTA GCA GCA CCG GAT TAA CAT AAC TTT CCA AAA TGT TGT ACG AAC CTT TAA ACA AAA
thr gln leu leu leu val gln leu his his phe leu ala ala pro asp OCH his asn phe pro lys cys cys thr asn arg och thr lys
3871/1291
ACA GTT CAC CTC CCT TTT CTA TAC TAT TGT CTG GCA GCA GTT GTT TGT TGT TAA AAA TAA CAG CCA TTG TAA TGA GAC GCA CAA ACT AAT
thr val his leu pro phe leu tyr tyr cys leu arg ala val val cys cys och lys och gln pro leu och OPA asp ala gln thr asn
3961/1321
ATC ACA AAC TGG AAA TGT CTA TCA TCA TAT AGT TGC TGA TAT CAT GGA GAT AAT TAA AAT GAT AAC CAT CTC OCA AAT AAA TAA GTA TTT
ile thr asn trp lys cys leu ser ile tyr ser cys OPA tyr his gly asp asn och asn asp asn his leu ala asn lys och val phe
4051/1351
TAC TOT TTT COT AAC AGT TTT GEA ATA AAA AAA CCT ATA AAT ATT CCG GAT TAT TCA TAC COT CCC ACC ATT GGG CGC GGA TGT ATT CTA
tyr cys phe arg asn ser phe val ile lys lys pro ile asn ile pro asp tyr ser tyr arg pro thr ile gly arg gly ser met leu
4141/1381
CTA GEA AAT CAG TCA CAC CAA GGC TTC AAT AAG GAA CAC ACA AGC AAG ATG GTA AGC GCT ATT GTT TTA TAT GTG CTT TTG GCG GCG GCG
leu val asn gln ser his gln gly phe asn lys glu his thr ser lys met val ser ala ile val leu tyr val leu leu ala ala
4231/1411
CGC CAT TCT GCC TTT CCG GCG GAT CTT gga tcc CAT CAT CAC CAC CAC CAC att gaa gga gga GAA TTC CAG GTG CAG CTG AAG GAG TCA
ala his ser ala phe ala ala asp leu gly ser his his his his his his ile glu gly arg glu phe gln val gln leu lys glu ser
4321/1441
GGA CTT GGC CTG GTG GCG CCC TCA CAG AGC CTG TCC ATC ACA TCC ACT GTC TCA GGG TTC TCA TTA ACC AGC TAT GGT GTA ACC TCG GTT
gly pro gly leu val ala pro ser gln ser leu ser ile thr cys thr val ser gly phe ser leu thr ser tyr gly val ser trp val
4411/1471
CGC CAG CTT CCA GGA AAG GGT CTG GAG TCG CTG GGA GTA ATA TGG GGT GAT GGG AGC ACA AAT TAT CAT TCA GCT CTC ACA TCC AGA CTG
arg gln pro pro gly lys gly leu glu trp CTG GGA GTA ATA TGG GGT GAT GGG AGC ACA AAT TAT CAT TCA GCT CTC ACA TCC AGA CTG
4501/1501
AGC ATC AGC AAG GAT AAC TCC AAG AGC CAA GTT TTC TTA AAA CTG AAC AGT CTG CAA ACT GAT GAC ACA GCC ACG TAC TAC TGT GCC AAA
ser ile ser lys asp asn ser lys ser gln val phe leu lys leu asn ser leu gln thr asp asp thr ala thr tyr tyr cys ala lys
4591/1531
AGG GGA GGC TAT GGT AAC TAC TAT GGT ATG GAC TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GCG GGA
arg gly gly tyr gly asn tyr tyr ala met asp tyr trp gly gln gly thr ser val thr val ser ser gly gly gly gly ser gly gly
4681/1561
GCT GGC TCT GGC GGT GGC GGA TCG GAC ATT GTG ATG ACC CAG TCT CAC AAA TTC ATG TCC ACA TCA GEA GGA GAC AGG GTC AGC ATC ACC
gly gly ser gly gly gly gly ser asp ile val met thr gln ser his lys phe met ser thr ser val gly asp arg val ser ile thr
4771/1591
TGC AAG GCC AGT CAG GAT GTG AGT ACT GCT GTA GCC TGG TAT CAA CAA AAA CCA GCG CAA TCT CCT AAA CTA CTG ATT TAC TGG GCA TCC
cys lys ala ser gln asp val ser thr ala val ala trp tyr gln gln lys pro gly gln ser pro lys leu leu ile tyr trp ala ser
4861/1621
ACT CCG CAC ACT GGA GTC CCT GAT CCG TTC ACA GGC AGT GGA TCT GGG ACA GAT TAT ACT CTC ACC ACC AGC AGT GTG CAG GCT GAA GAC
thr arg his thr gly val pro asp arg phe thr gly ser gly ser gly thr asp tyr thr leu thr ile ser ser val gln ala gly asp
4951/1651
CTG GCA CTT TAT TAC TGT CAG CAA CAT TAT AGC ACT CCT CCG AGC TTC GGT GGA GGC ACC AAG CTG GAA ATC AAA CCG OCT CCC GCG GGA
leu ala leu tyr tyr cys gln gln his tyr ser thr pro pro thr phe gly gly gly thr lys leu glu ile lys arg ala pro gly gly

CDR des régions variable et VL

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fin ScFv 2 gly

pACgp67-ScFv461 -> 1-phase Translation

Fig. 1C

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[illegible]

pACgp67-ScFv461 -> 1-phase Translation

Fig. 1D

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7651/2551 7681/2561 7711/2571
 GAT TTG CAC AAG CAC AAT TTC ATA CAC AAC GAC ATA AAA CTC GAA AAT GTC TGA TAT TTC GAA GCA CTT GAT CGC GTG TAT GTT TGC GAT
 asp leu his lys his asn phe ile his asn asp ile lys leu glu asn val leu tyr phe glu ala leu asp arg val tyr val cys asp
 7741/2581 7771/2591
 TAC GGA TTG TGC AAA CAC GAA AAC TCA CTT AGC GTG CAC GAC GGC ACG TTG GAG TAT TTT AGT CCG GAA AAA ATT CGA CAC ACA ACT ACT
 tyr gly leu cys lys his glu asn ser leu ser val his asp gly thr leu glu tyr phe ser pro glu lys ile arg his thr thr met
 7831/2611 7861/2621
 CAC GTT TCG TTT GAC TGG TAC CGC GCG TGT TAA CAT ACA AGT TGC TAA CCG GCG GGT GGT AAT CAT GGT CAT AGC TGT TTC CTG TGT GAA
 his val ser phe asp trp tyr ala ala cys OCH his thr ser cys OCH pro ala val arg asn his gly his ser cys phe leu cys glu
 7921/2641 7951/2651
 ATT GTT ATC CGC TCA CAA TTC CAC ACA ACA TAC GAG CCG GAA GCA TAA AGT GTA AAG CCT GGG GTG CCT AAT GAG TGA GCT AAC TCA CAT
 ile val ile arg ser gln phe his thr thr tyr glu pro glu ala OCH ser val lys pro gly val pro asn glu OPA ala asn ser his
 8011/2671 8041/2681
 TAA TTG CGT TGC GCT CAC TGC CCG CTT TCC AGT CCG GAA ACC TGT CGT GGC AGC TGC ATT AAT GAA TCG GCC AAC GCG CCG GGA GAG GCG
 OCH leu arg cys ala his cys pro leu ser ser arg glu thr cys arg ala ser cys ile asn glu ser ala asn ala arg gly glu ala
 8101/2701 8131/2721
 GTT TGC GTA TTG GGC GCT CTT CCG CTT CCT CGC TCA CTG ACT CGC TGC GCT CCG TCG TTC GGC TCG GGC GAG CCG TAT CAG CTC ACT CAA
 val cys val leu gly ala leu pro leu pro arg ser leu thr arg cys ala arg ser phe gly cys gly glu arg tyr gln leu thr gln
 8191/2731 8221/2741
 AGG CCG TAA TAC GGT TAT CCA CAG AAT CAG GGG ATA ACG CAG GAA AGA ACA TGT GAG CAA AAG GCG AGC AAA AGG CCA GGA ACC GTA AAA
 arg arg OCH tyr gly tyr pro gln asn gln gly ile thr gln glu arg thr cys glu gln lys ala ser lys arg pro gly thr val lys
 8281/2761 8311/2771
 AGG CCG CGT TGC TGG CGT TTT TCC ATA GGC TCC GCG CCC CTG ACG AGC ATC ACA AAA ATC GAC GCT CAA GTC AGA GGT GGC GAA ACC CGA
 arg pro arg cys trp arg phe ser ile gly ser ala pro leu thr ser ile thr lys ile asp ala gln val arg gly gly glu thr arg
 8371/2791 8401/2801
 CAG GAC TAT AAA GAT ACC AGG CGT TTC CCC CTG GAA GCT CCC TCG TGC GCT CTC CTG TTC CGA CCC TGC CGC TTA CCG GAT ACC TGT CCG
 gln asp tyr lys asp thr arg arg phe pro leu glu ala pro ser cys ala leu leu phe arg pro cys arg leu pro asp thr cys pro
 8461/2821 8491/2831
 CCT TTC TCC CTT CCG GAA GCG TGG CCG TTT CTC ATA GGT CAC GGT GTA GGT ATC TCA GTT CCG TGT AGG TCG TTC GCT CCA AGC TCG OCT
 pro phe ser leu arg glu ala trp arg phe leu ile ala his ala val gly ile ser val arg cys arg ser phe ala pro ser trp ala
 8551/2851 8581/2861
 GTG TGC ACG AAC CCC CCG TTC AGC CCG ACC GCT GCG CCT TAT CCG GTA ACT ACT GTC TTG AGT CCA ACC CCG TAA GAC ACG ACT TAT CCG
 val cys thr asp pro pro phe ser pro thr ala ala pro tyr pro val thr ile val leu ser pro thr arg OCH asp thr thr tyr arg
 8641/2881 8671/2891
 CAC TGG CAG CAG CCA CTG GTA ACA GGA TTA GCA GAG CGA GGT ATG TAG GCG GTG CTA CAG ACT TCT TGA AGT GGT GGC CTA ACT ACG GCT
 his trp gln gln pro leu val thr gly leu ala glu arg gly met AMB ala val leu gln ser ser OPA ser gly gly leu thr thr ala
 8731/2911 8761/2921
 ACA CTA GAA GGA CAG TAT TTG GTA TCT GCG CTC TCT TGA AGC CAG TTA CCT TCG GAA AAA GAG TTG GTA GCT CTT GAT CCG GCA AAC AAA
 thr leu glu gly gln tyr leu val ser ala leu cys OPA ser gln leu pro ser glu lys glu leu val ala leu asp pro ala asn lys
 8821/2941 8851/2951
 CCA CCG CTG GTA GCG GTG GTT TTT TTG TTT OCA AGC AGC AGA TTA CCG CCA GAA AAA AAG GAT CTC AAG AAG ATC CTT TGA TCT TTT CTA
 pro pro leu val ala val val phe leu phe ala ser ser arg leu arg ala glu lys lys asp leu lys lys ile leu OPA ser phe leu
 8911/2971 8941/2981
 CCG GGT CTG ACG CTC AGT GGA AGC AAA ACT CAC GTT AAG GGA TTT TCG TCA TGA GAT TAT CAA AAA GGA TCT TCA CCT AGA TCC CTT TAA
 arg gly leu thr leu ser gly thr lys thr his val lys gly phe trp ser OPA asp tyr gln lys gly ser ser pro arg ser phe OCH
 9001/3001 9031/3011
 ATT AAA AAT GAA GTT TTA AAT CAA TCT AAA GTA TAT ATG AGT AAA CTT GGT CTG ACA GTT ACC AAT GCT TAA TCA GTG AGG CAC CTA TCT
 ile lys asn glu val leu asn gln ser lys val tyr met ser lys leu gly leu thr val thr asn ala OCH ser val arg his leu ser
 9091/3031 9121/3041
 CAG CGA TCT GTC TAT TTC GTT CAT CCA TAG TTG CTT GAC TCC CCG TCG TGT AGA TAA CTA CGA TAC GCG AGG OCT TAC CAT CTG GCG CCA
 gln arg ser val tyr phe val his pro AMB leu pro asp ser pro ser cys arg OCH leu arg tyr gly arg ala tyr his leu ala pro
 9181/3061 9211/3071
 GTG CTG CAA TGA TAC CCG GAG ACC CAC GCT CAC CCG CTC CAG ATT TAT CAG CAA TAA ACC AGC CAG CCG GAA GCG CCG AGC GCA GAA GTG
 val leu gln OPA tyr arg glu thr his ala his arg thr gln ile tyr gln gln OCH thr ser gln pro glu gly pro ser ala glu val
 9271/3091 9301/3101
 GTC CTG CAA CTT TAT CCG CCT CCA TCC AGT CTA TTA ATT GTT GCG GCG AAG CTA GAG TAA GTA GTT CCG CAG TTA ATA GTT TCC GCA ACG
 val leu gln leu tyr pro pro pro ser ser leu leu ile val ala gly lys leu glu OCH val val arg gln leu ile val cys ala thr
 9361/3121 9391/3131
 TTG TTG CCA TTG CTA CAG GCA TCG TGG TGT CAC GCT CGT COT TTG GTA TGG CTT CAT TCA GCT CCG GTT CCC AAC GAT CAA GCG GAG TTA
 leu leu pro leu leu gln ala ser trp cys his ala arg arg leu val trp leu his ser ala pro val pro asn asp gln gly glu leu
 9451/3151 9481/3161
 CAT GAT CCC CCA TGT TGT GCA AAA AAG CCG TTA OCT CCT TCG GTC CTC CGA TCG TTG TCA GAA GTA AGT TGG CCG CAG TGT TAT CAC TCA
 his asp pro pro cys cys ala lys lys arg leu ala pro ser val leu arg ser leu ser glu val ser trp pro gln cys tyr his ser
 9541/3181 9571/3191
 TCG TTA TGG CAG CAC TGC ATA ATT CTC TTA CTG TCA TGC CAT CCG TAA GAC GCT TTT CTG TGA CTG GTG AGT ACT CAA CCA AGT CAT TCT
 trp leu trp gln his cys ile ile leu leu leu ser cys his pro OCH asp ala phe leu OPA leu val ser thr gln pro ser his ser
 9631/3211 9661/3221
 GAG AAT AGT GTA TGC GGC GAC CGA GTT GCT CTT GCG CCG COT CAA TAC GCG ATA ATA CCG CCG CAC ATA GCA GAA CTT TAA AAG TGC TCA
 glu asn ser val cys gly asp arg val ala leu ala arg arg gln tyr gly ile ile pro arg his ile ala glu leu OCH lys cys ser
 9721/3241 9751/3251
 TCA TTG GAA AAC GTT CTT CCG GGC GAA AAC TCT CAA GGA TCT TAC CCG TGT TGA GAT CCA GTT CGA TGT AAC CCA CTC GTG CAC CCA ACT
 ser leu glu asn val leu arg gly glu asn ser gln gly ser tyr arg cys OPA asp pro val arg cys asn pro leu val his pro thr
 9811/3271 9841/3281
 GAT CTT CAG CAT CTT TTA CTT TCA CCA GCG TTT CTG GGT GAG CAA AAA CAG GAA GGC AAA ATG CCG CAA AAA AGG GAA TAA GCG CGA CAC
 asp leu gln his leu leu leu ser pro ala phe leu gly glu gln lys gln glu gly lys met pro gln lys arg glu OCH gly arg his
 9901/3301 9931/3311
 GGA AAT GTT GAA TAC TCA TAC TCT TCC TTT TTC AAT ATT ATT GAA GCA TTT ATC AGG GTT ATT GTC TCA TGA GCG GAT ACA TAT TTG AAT
 gly asn val glu tyr ser tyr ser ser phe phe asn ile ile glu ala phe ile arg val ile val ser OPA ala asp thr tyr leu asn
 9991/3331 10021/3341
 GTA TTT AGA AAA ATA AAC AAA TAG GGG TTC CCG CCA CAT TTC CCC GAA AAG TGC CAC CTG ACG TCT AAG AAA CCA TTA TTA TCA TGA CAT
 val phe arg lys ile asn lys AMB gly phe arg ala his phe pro glu lys cys his leu thr ser lys lys pro leu leu ser OPA his
 10081/3361 10111/3371
 TAA CCT ACA AAA ATA GGC GTA TCA CGA GGC CCT TTC GTC TCG CCG GGT TCG GTG ATG ACG GTG AAA ACC TCT GAC ACA TGC AGC TTC CCG
 OCH pro ile lys ile gly val ser arg gly pro phe val ser arg val ser val met thr val lys thr ser asp thr cys ser ser arg
 10171/3391 10201/3401
 AGA CCG TCA CAG CTT GTC TGT AAG CCG ATG CCG GGA GCA GAC AAG CCC GTC AGG GCG COT CAG CCG GTG TTG GCG GGT GTC GCG GCT GCG
 arg arg ser gln leu val cys lys arg met pro gly ala asp lys pro val arg ala arg gln arg val leu ala gly val gly ala gly

Fig 1E

pACgp67-ScFv461 -> 1-phase Translation

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10261/3421 10291/3431 10321/3441
 TTA ACT ATG CGG CAT CAG AGC AGA TTG TAC TGA GAG TGC ACC ATA TGC GGT GTG AAA TAC CGC ACA GAT GCG TAA GGA GAA AAT ACC GCA
 leu thr met arg his gln ser arg leu tyr CPA glu cys thr ile cys gly val lys tyr arg thr asp ala CCH gly glu asp thr ala
 10351/3451 10381/3461 10411/3471
 TCA GGC GCC ATT CGC CAT TCA GGC TGC GCA ACT GTT GGG AAG GGC GAT CGG TGC GGG CTT CTT CGC TAT TAC GGC AGC TGG CGA AAG GCG
 ser gly ala ile arg his ser gly cys ala thr val gly lys gly asp arg cys gly pro leu arg tyr tyr ala ser trp arg lys gly
 10441/3481 10471/3491 10501/3501
 GAT GTG CTG CAA GGC GAT TAA GTT GGG TAA CGC CAG GGT TTT CCC AAT CAC GAC GTT GTA AAA CGA CCG CCA GTG CC
 asp val leu gln gly asp OCH val gly OCH arg gln gly phe pro ser his asp val val lys arg arg pro val

pACgp67-ScFv350 -> 1-phase Translation

Fig. 2B

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2431/811 AAT TAA ATA GCT TGC GAC GCA ACG TGC ACG ATC TGT GCA CCG GTT CCG GCA CGA GCT TTG ATT GEA ATA AGT TTT TAC GAA GCG ACG ACA
asn och ile ala cys asp ala thr cys thr ile cys ala arg val pro ala arg ala leu ile val ile ser phe tyr glu ala met thr
2521/841 TGA CCC CCG TAG TGA CAA CGA TCA CCG CCA AAA GAA CTG CCG ACT ACA AAA TTA CCG AGT ATG TCG GTG ACG TTA AAA CTA TCA AGC CAT
OPA pro pro AMB OPA gln arg ser arg pro lys glu leu pro thr thr lys leu pro ser met ser val thr leu lys leu leu ser his
2611/871 CCA ATC GAC CGT TAG TCG AAT CAG GAC CCG TGG TGC GAG AAG CCG CGA AGT ATG GCG AAT GGA TCG TAT AAC GTG TGG AGT CCG CTC ATT
pro ile asp arg AMB ser asn gln asp arg trp cys glu lys pro arg ser met ala asn ala ser tyr asn val trp ser pro leu ile
2701/901 AGA GCG TCA TGT TTA GAC AAG AAA GCT ACA TAT TTA ATT GAT CCC GAT GAT TTT ACT GAT AAA TTG ACC CTA ACT CCA TAC ACG GTA TTC
arg ala ser cys leu asp lys lys ala thr tyr leu ile asp pro asp asp phe ile asp lys leu thr leu thr pro tyr thr val phe
2791/931 TAC AAT GGC GGG GTT TTG GTC AAA ATT TCC GGA CTG CGA TTG TAC ATG CTG TTA ACG GCT CCG CCC ACT ATT AAT GAA ATT AAA AAT TCC
tyr asn gly gly val leu val lys ile ser gly leu arg leu tyr met leu leu thr ala pro pro thr ile asn glu ile lys asn ser
2881/961 AAT TTT AAA AAA CCG ACG AAG AGA AAC ATT TGT ATG AAA GAA TCC GTA GAA GGA AAG AAA AAT GTC GTC GAC ATG CTG AAC AAC AAG ATT
asn phe lys lys arg ser lys arg asn ile cys met lys glu cys val glu gly lys lys asn val val asp met leu asn asn lys ile
2971/991 AAT ATG CCT CCG TGT ACA AAA AAA ATA TTG AAC GAT TTG AAA GAA AAC AAT GTA CCG CCG GCG GGT ATG TAC AGG AAG AGG TTT ATA CTA
asn met pro pro cys ile lys lys ile leu asn asp leu lys glu asn asn val pro arg gly gly met tyr arg lys arg phe ile leu
3061/1021 AAC TGT TAC ATT GCA AAC GTG GTT TCG TOT GCC AAG TOT GAA AAC CGA TOT TTA ATC AAG GCT CTG ACG CAT TTC TAC AAC CAC GAC TCC
asn cys tyr ile ala asn val val ser cys ala lys cys glu asn arg cys leu ile lys ala leu thr his phe tyr asn his asp ser
3151/1051 AAG TGT GTG GGT GAA GTC ATG CAT CTT TTA ATC AAA TCC CAA GAT GTG TAT AAA CCA CCA AAC TGC CAA AAA ATG AAA ACT GTC GAC AAG
lys cys val gly glu val met his leu leu ile lys ser gls asp val tyr lys pro pro asn cys gln lys met lys thr val asp lys
3241/1081 CTC TGT CCG TTT GCT GGC AAC TGC AAG GGT CTC AAT CCT ACT TGT AAT TAT TGA ACA ATA AAA CAA TTA TAA ATG CTA AAT TTG TTT TTT
leu cys pro phe ala gly asn cys lys gly leu asn pro ile cys asn tyr OPA ile ile lys gln leu och met leu asn leu phe phe
3331/1111 ATT AAC GAT ACA AAC CAA ACG CAA CAA GAA CAT TTG TAG TAT TAT CTA TAA TTG AAA ACG CGT AGT TAT AAT CCG TGA GGT AAT ATT TAA
ile asn asp thr asn gln thr gln gln glu his leu AMB tyr tyr leu och leu lys thr arg ser tyr asn arg OPA gly asn ile och
3421/1141 AAT CAT TTT CAA ATG ATT CAC AGT TAA TTT GCG ACA ATA TAA TTT TAT TTT CAC ATA AAC TAG ACG CCT TGT CCG CTT CTT CTT COT ACT
asn his phe gln met ile his ser och phe ala thr ile och phe tyr phe his ile asn AMB thr pro cys arg leu leu leu arg ile
3511/1171 CCT TGT CTT TTT CAT TTT TCT CCT CAT AAA AAT TAA CAT AGT TAT TAT COT ATC CAT ATA TGT ATC TAT COT ATA GAG TAA ATT TTT TGT
pro ser leu phe his phe ser pro his lys asn och his ser tyr tyr arg ile his ile cys ile tyr arg ile glu och ile phe cys
3601/1201 TGT CAT AAA TAT ATA TOT CTT TTT TAA TCG GGT GTA TAG TAC CCG TGC GCA TAG TTT TTC TGT AAT TTA CAA CAG TCC TAT TTT CTG GEA
cys his lys tyr ile cys leu phe och trp gly val AMB tyr arg cys ala AMB phe phe cys asn leu gln gls cys tyr phe leu val
3691/1231 GTT CTT CCG AGT GTG TTG CTT TAA TTA TTA AAT TTA TAT AAT CAA TGA ATT TCG GAT COT CCG TTT TGT ACA ACA TOT TCC CCG CAT AGT
val leu arg ser val leu leu och leu leu asn leu tyr asn gln OPA ile trp asp arg arg phe cys thr ile cys cys arg his ser
3781/1261 ACC GAG CTT CTT CTA GTT CAA TTA CAC CAT TTT TTA GCA GCA CCG GAT TAA CAT AAC TTT CCA AAA TOT TGT AGC AAG COT TAA ACA AAA
thr gln leu leu leu val gln leu his his phe leu ala ala pro asp och his asn phe pro lys cys cys thr asn arg och thr lys
3871/1291 ACA GTT CAT CTC CTT TTT TTA TAC TAT TOT CTG TGA GCA GTT GTT TOT TAT TAA AAA TAA CAG CCA TTG TAA TGA GAC GCA CAA ACT AAT
thr val his leu pro phe leu tyr tyr cys leu arg ala val val cys cys och lys och CAG gln pro leu och OPA asp ala gln thr asn
3961/1321 ACC ACA AAC TGG AAA TGT CTA TCA ATA TAT AGT TGC TGA TAC CAT GGA GAT AAT TAA AAT GAT AAC CAT CTC GCA AAT AAA TAA GTA TTT
ile thr asn trp lys cys leu ser ile tyr ser cys OPA tyr his gly asp asn och asn asp asn his leu ala asn lys och val phe
4051/1351 TAC TGT TTT COT AAC AGT TTT GEA ATA AAA AAA CTT ATA AAT ATC CCG GAT TAT TCA TAC COT CCC ACC ATC GCG CCG GGA TGT ATT CTA
tyr cys phe arg asn ser phe val ile lys lys pro ile asn ile pro asp tyr ser tyr arg pro thr ile gly arg gly ser met leu
4141/1381 CTA GCA AAT CAG TCA CAC CAA GGC TTC AAT AAG GAA CAC ACA AGC AAG ATG GTA AGC GCT ACT GTT TTA TAT GTG CTT TTG GCG GCG GCG
leu val asn gln ser his gln gly phe asn lys glu his thr ser lys met val ser ala ile val leu tyr val leu leu ala ala ala
4231/1411 CCG CAT TGT GCG TTT GCG CCG GAT CTT GGA TCC CAT CAT CAC CAC CAC CAC att gaa gga aga GAA TTT CAG GTG CAA CTG CAG CAG TCT
ala his ser ala phe ala ala asp leu gly ser his his his his his his ile glu gly arg glu phe gln val gln leu gln gln ser
4321/1441 GGG GCT GAA CTG GCA AAA CCT GCG GCC TCA GTG AAG CTG TCC TGC AAG GCT TCT GGC CAC ACC TTT ACT AGC TAC TGG ATG CAC TGG GTA
gly ala glu leu ala lys pro gly ala ser val lys leu ser cys lys ala ser gly his thr phe thr ser tyr trp met his trp val
4411/1471 AAA CAG AGG CCT GGA CAG GGT CTG GAA TGG ATT GGA TAC ATT AAT CTT AGC AGT GGT TAT ATT AAG TAC AAT CAG GAG TTC AAG GAC AAG
lys gln arg pro gly gln gly leu glu trp ile gly tyr ile asn leu ser ser gly tyr ile lys tyr asn gln glu phe lys asp lys
4501/1501 GCC ACA TTG ACT GCA GAC AAA TCC TCC AAC ACA GCT TAC ATG CAT CTG AGC ACC CTG ACA TAT GAG GAC TCT GCA GTC TAT TAC TGT GCA
ala thr leu thr ala asp lys ser ser asn thr ala tyr met his leu ser ser leu thr tyr glu asp ser ala val tyr tyr cys ala
4591/1531 AGG GCA GGT CAG GCT ACG ACC TTT GAC TAC TGG TGC CCA GGC ACC ACT CTC ACA GTC TCC TCA GGT GGA GGC GGT GCA GGT GCG
arg ala ala gln ala thr thr phe asp tyr trp gly gln gly thr thr leu thr val ser ser gly gly gly gly ser gly gly gly gly
4681/1561 TCT GCG GGT GCG GGA TCG GAC ATT GTG ATG ATC CAG TCT CAC AAA TTC ATG TCC ACA TCA TCA GGA GAC AGG GTC AGC ATC ACC TGC AAG
ser gly gly gly gly ser asp ile val met ile gln ser his lys phe met ser thr ser val gly asp arg val ser ile thr cys lys
4771/1591 GCC AGT CAG GAT GTG AGT ACT GCT GEA GCC TGG TAT CAA CAA AAA CCA GCG CAA TCT CTT AAA CTA CTG ATT TAC TGG GCA TCC ACC CCG
ala ser gln asp val ser thr ala val ala trp tyr gln gln lys pro gly gln ser pro lys leu leu ile tyr trp ala ser thr arg
4861/1621 CAC ACT GGA GTC CTT GAT CCG TTC ACA GGC AGT GGA TCT GCG ACA GAT TAT ACT CTC ACC ATC AGC AGT GTG CAG GCT GAA GAC CTG GCA
his thr gly val pro asp arg phe thr gly ser gly ser gly thr asp tyr thr leu thr ile ser ser val gln ala glu asp leu ala
4951/1651 CTT TAT TAC TGT CAG CAA CAT TAT AGC ACT CTT CCG ACG TTC GGT GGA GGC ACC AAG CTG GAA ATT AAA CCG GCT CTT GCG GGA GGT TAA
leu tyr tyr cys gln gln his tyr ser thr pro pro thr phe gly gly gly thr lys leu glu ile lys arg ala pro pro gly cys och

: CDR des régions variables VH et V

7/21

fin scfv

2gly 1Cys

Stop

pap tide de fonction

Fig. 2C

pACgp67-ScFv350 -> 1-phase Translation

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5041/1681
 aga tct gat cct TTC CTG GGA CCC GGC AAG
 arg ser asp pro phe leu gly pro gly lys
 5131/1711
 GTC GTT GGA TGG AAA GGA AAA GAG TTC TAC
 val val gly trp lys gly lys glu phe tyr
 5221/1741
 ATG GAT GTT TTC CTT GTT GTC AAC ATG COT
 met asp val phe leu val val asn met arg
 5311/1771
 GAC TAT GTA CCT CAT GAC GTG ATT AGG ATC
 asp tyr val pro his asp val ile arg ile
 5401/1801
 GGC GGC TGC CCA ATA ATG AAC CTT CAC TCT
 gly gly cys pro ile met asn leu his ser
 5491/1831
 CCC ATC GTT TAC ATC GGT ACC GAC TCT GCT
 pro ile val tyr ile gly thr asp ser ala
 5581/1861
 GAC GCA CCT CTG TTC ACT GGT CCG GCG TAT
 asp ala pro leu phe thr gly pro ala tyr
 5671/1891
 TTT ACA GAC AAT TGT TGT AGG TAT TTT AAT
 phe thr asp asn cys cys thr tyr phe asn
 5761/1921
 GCG TCT TTA TAC CTG AAT TEA AAT ATT AAA
 ala ser leu tyr leu asn leu asn ile lys
 5851/1951
 CCG ATG GCT GGA CTA TCT AAT GGA TTT TCG
 pro met ala gly leu ser asn gly phe ser
 5941/1981
 TGT GTT TTG TTT TGT AAT AAA GGT TCG ACG
 cys val leu phe cys asn lys gly ser thr
 6031/2011
 CTC GAC GTA AAC AGC TTA AAT AAA GCT TGG
 leu asp val asn thr leu asn lys ala trp
 6121/2041
 TCG TCG TTA GAA GTT GCT TCC GAA GAC GAT
 ser ser leu glu val ala ser glu asp asp
 6211/2071
 GTA GTT GAG CTT TTT GGA ATT ATT TCT GAT
 val val glu leu phe gly ile ile ser asp
 6301/2101
 TTA GAA AGC GAT GGT GCA GGC GGT GGT AAC
 leu glu ser asp gly ala gly gly gly asn
 6391/2131
 GGT GGA GGC GCA GGC GGG GCT GGC GGC GGA
 gly gly gly ala gly gly ala gly gly gly
 6481/2161
 GGC AAC ACA GTC GGC ACC TCA ACT ATT GTA
 gly asn thr val gly thr ser thr ile val
 6571/2191
 CTA ATA GCT TCC AAC AAT TGT TGT CTG TCG
 leu ile ala ser asn asn cys cys leu ser
 6661/2221
 ATC GAT GGT GGT GGT GGT GGT GGA GGC GCT
 ile asp gly gly gly gly gly gly gly gly
 6751/2251
 TTA GTT TGT TCG GCG ACT ATT GTG GGC ACC
 leu val cys ser arg thr ile val gly thr
 6841/2281
 GGT GGC AAT TCA ATA TTA TAA TTG GAA TAC
 gly gly asn ser ile leu OCH leu glu tyr
 6931/2311
 AAC AAC CCG TCA ATG CAA GCA ATT GGA TTG
 asn asn arg ser met OCH ala ile val leu
 7021/2341
 TTG TAG TGG CGA GAC ACT TCG CTG TCG TCG
 leu X3 trp arg asp thr ser leu ser ser
 7111/2371
 ACA TCT CTG TTC ACC ACC ACT GTG TTG TCG
 thr ser leu phe ser thr thr val leu ser
 7201/2401
 GCA TCA ATT TTG TTG TTC CTA TTA TTG AAT
 ala ser ile leu leu phe leu leu leu asn
 7291/2431
 CAA ACG CTG GGA CAA TTT TAC GAA AAC TGC
 gln thr leu val gln phe tyr glu asn cys
 7381/2461
 TCG CAC AAC CCC ACT AGC AAA TTG TAC TTG
 ser his lys pro thr ser lys leu tyr leu
 7471/2491
 AGC GAC CAC CCA AAT TTT ATA AAA ATC TAT
 ser asp his pro asn phe ile lys ile tyr
 7561/2521
 TTA TTT GAA ACA CTA CAA ATT AAA GGC GAG
 leu phe glu thr leu gln ile lys gly glu
 5071/1691
 AAC CAA AAA CTC ACT CTC TTC AAG GAA ATC
 asn gln lys leu thr leu phe lys glu ile
 5161/1721
 AGG GAA ACT TGG ACC CCG TTC ATG GAA GAC
 arg glu thr trp thr arg phe met glu asp
 5251/1751
 CCC ACT AGA CCC AAC CGT TGT TAC AAA TTC
 pro thr arg pro asn arg cys tyr lys phe
 5341/1781
 GTC GAG CCT TCA TGG GTG GGC AGC AAC AAC
 val glu pro ser trp val gly ser asn asn
 5431/1811
 GAG TAC ACC AAC TCG TTC GAA CAG TTC ATC
 glu tyr thr asn ser phe glu gln phe ile
 5521/1841
 GAA GAG GAG GAA ATT CTC CTT GAA GTT TCC
 glu glu glu glu ile leu leu glu val ser
 5611/1871
 TAA AAC ACG ATA CAT TGT TAT TAG TAC ATT
 OCH asn thr ile his cys tyr AMS tyr ile
 5701/1901
 AAT TCA TTA AAT TTA TAA TCT TTA GCG TGG
 asn ser leu asn leu OCH ser leu gly trp
 5791/1931
 TCC TCA ATA GAT TTG TAA AAT AGG TTT CGA
 ser ser ile asp leu OCH asn arg phe arg
 5881/1961
 CTC AAC GCC ACA AAA CTT GCC AAA TCT TGT
 leu asn ala thr lys leu ala lys ser cys
 5971/1991
 TCG TTC AAA ATA TEA TGC GCT TTT GTA TTT
 ser phe lys ile leu cys ala phe val phe
 6061/2021
 ACA TAT TTA ACA TCG GGC GTG TTA GCT TTA
 thr tyr leu thr ser gly val leu ala leu
 6151/2051
 TTT GCC ATA GCC ACA CGA CCG CTA TTA ATT
 phe ala ile ala thr arg arg leu leu ile
 6241/2081
 TCG GCG CGT TTT TGG GCG GGT TTC AAT CTA
 cys gly arg phe trp ala gly phe asn leu
 6331/2111
 ATT TCA GAC GGC AAA TCT ACT AAT GCG GCG
 ile ser asp gly lys ser thr asn gly gly
 6421/2141
 GCG GGA GCG GCG GGT GGT GCG GGT GAC GCA
 gly gly gly gly gly gly gly gly asp ala
 6511/2171
 CTG GTT TCG GCG GCG GTT TTT GGT TTG ACG
 leu val ser gly ala val phe gly leu thr
 6601/2201
 TCT AAA GGT GCA GCG GGT TGA GGT TCC GTC
 ser lys gly ala ala gly OPA gly ser val
 6691/2231
 GGA ATG TTA GCG ACG GGA GAA GGT GGT GCG
 gly met leu gly thr gly glu gly gly gly
 6781/2261
 GGC GCA GGC GCG GCT GCG TCC ACA ACG GAA
 gly ala gly ala ala gly cys thr thr glu
 6871/2291
 TAA AGA GAT TGT CTC AAG CTC CCG ACG CCG
 OCH arg asp cys leu lys leu arg thr pro
 6961/2321
 TCG TAC ATG TAT GGT TTG TTG TCA AAA ACG
 thr tyr met tyr ala leu leu ser lys thr
 7051/2351
 TAA ATG TTG TTT TTG ACA AAT TCC GCT TCC
 OCH met leu phe leu ile ile cys ala ser
 7141/2381
 AAA CAA GAT TGT ACA GAT TCA TAT CTA CGA
 lys ser OCH lys ser ala ile ser ile val
 7231/2411
 TAA AGA GAT TGT CTC AAG CTC CCG ACG CCG
 OCH arg asp cys leu lys leu arg thr pro
 7321/2441
 AAA AAC GTC AAA ACT CCG TAT AAA ATA ATC
 lys asn val lys thr arg tyr lys ile ile
 7411/2471
 CAG AAA ACA AST TCG CCG CAC AAT TTT AAC
 gln lys thr ile ser ala his asn phe asn
 7501/2501
 CTT AAT CAC GGT TCC ACT AAC AAC CAA GTG
 phe asn his gly ser ile asn asn gln val
 7591/2531
 CTT TCG TAC CAA CTT GTT ACC AAT ATT ATT
 thr thr thr gln leu val ser asn ile ile
 5101/1701
 CCG AAT GGT AAA CCC GAC ACG ATG AAG CTT
 arg asn val lys pro asp thr met lys leu
 5191/1731
 AGC TTC CCG ATT GTT AAC GAC CAA GAA GTG
 ser phe pro ile val asn asp gln glu val
 5281/1761
 CCG GCC CAA CAC GCT CTG COT TGC GAC CCC
 leu ala gln his ala leu arg cys asp pro
 5371/1791
 GAG TAC CCG ATT AGC CTG COT AAG AAG GCG
 glu tyr arg ile ser leu ala lys lys gly
 5461/1821
 GAT CCG GTC ATC TGG GAG AAC TTC TAC AAG
 asp arg val ile trp glu asn phe tyr lys
 5551/1851
 CTG GTG TTC AAA GGA AAC GAG TTT GCA CCA
 leu val phe lys val lys glu phe ala pro
 5641/1881
 TAT TAA GCG CTA GAT TCT GTG COT TGT TGA
 tyr OCH ala leu asp ser val arg cys OFA
 5731/1911
 TAT GTT AGA GCG AAA ATC AAA TGA TTT TCA
 tyr val arg ala lys ile lys OFA phe ser
 5821/1941
 TTA GTT TCA AAC AAG GGT TGT TTT TCC GAA
 leu val ser asn lys gly cys phe ser glu
 5911/1971
 AGC AGC AAT CTA GCT TTG TCG AAT TTC GTT
 ser ser asn leu ala leu ser ile phe val
 6001/2001
 CTT TCA TCA CTG TCG TTA GTG TAC AAT TGA
 leu ser ser leu ser leu val tyr asn OFA
 6091/2031
 TTA GCG CGA TTA CCG TCG TCG TCC CAA CCC
 leu gly arg leu ser ser ser ser gln pro
 6181/2061
 GTG TCG GCT AAC ACG TCC GCG ATC AAA TTT
 val ser ala asn thr ser ala ile lys phe
 6271/2091
 ACT GTG CCC GAT TTT AAT TCA GAC AAC ACG
 thr val pro asp phe asn ser asp asp thr
 6361/2121
 COT GGT GGA GCT GAT GAT AAA TCT ACC ATC
 gly gly gly ala asp asp lys ser thr ile
 6451/2151
 GAC CCG GGT TTA GCG TCA AAT GTC TCT TTA
 asp gly gly leu gly ser asn val ser leu
 6541/2181
 GGT CTG AGA CGA GTG CGA TTT TTT TCG TTT
 gly leu arg arg val arg phe phe ser phe
 6631/2211
 GGC ATT GGT GGA GCG GCG GCG AAT TCA GAC
 gly ile gly gly ala gly gly gly asn asp
 6721/2241
 GCG GGT GCG GCT GGT ATA ATT TGT TCT GGT
 gly gly ala ala gly ile ile cys ser gly
 6811/2271
 GGT GGT CTG CTT CGA GCG AGC GCT TCG GGT
 gly arg leu leu arg gly ser ala trp gly
 6901/2301
 ATT TCG CTA TCG TTT ACC GTG CCG ATA TTT
 ile ser leu ser phe thr val pro ile phe
 6991/2331
 ATA ACA AGC CTT TTC ATT TTT ACT ACA GCA
 ile thr ser leu phe ile phe thr thr ala
 7081/2361
 TCG TTG GCA AGC TTT AAA ATA TTT AAA AGA
 ser leu ala ser phe lys ile phe lys arg
 7171/2391
 GCA GTA TCG ACA GGT TCA AAA AAT TGA TGC
 ala val ser thr arg ser lys asn OPA cys
 7261/2421
 TTC GTC ATG GCC ACC ACA AAT GCT ACG CTG
 phe val met ala thr thr asn ala thr leu
 7351/2451
 AAC GCG CCG TTT GCG AAA ATA TCT ATT TTA
 asn gly arg phe gly lys ile ser ile leu
 7441/2481
 GCT GAC GAA ATA AAA GTT CAC CAG TTA ATG
 ala asp glu ile lys val his gln leu met
 7531/2511
 ATC GTG ATG GAC TAC ATT GAC TGT CCC GAT
 ile val met asp tyr ile asp cys pro asp
 7621/2541
 AGA CAG CTG TGT GAA GCG CTC AAC GAT TTG
 arg gln leu cys glu ala leu asn asp leu

pACgp67-ScFv350 -> 1-phase Translation

Fig 2D

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7551/2551
 CAC AAG CAC AAT TTC ATA CAC AAC GAC ATA
 his lys his asn phe ile his asn asp ile
 7741/2581
 TTG TGC AAA CAC GAA AAC TCA CTT AGC GTG
 leu cys lys his glu asn ser leu ser val
 7831/2611
 TCG TTT GAC TGG TAC GCG GCG TGT TAA CAT
 ser phe asp trp tyr ala ala cys OCH his
 7921/2641
 ATC GCG TCA CAA TTC CAC ACA ACA TAC GAG
 ile arg ser gln phe his thr thr tyr glu
 8011/2671
 COT TGC GCT CAC TGC CCG CTT TCC AGT CCG
 arg cys ala his cys pro leu ser ser arg
 8101/2701
 GTA TTG GGC GCT CTT CCG CTT CTT CCG TCA
 val leu gly ala leu pro leu pro arg ser
 8191/2731
 TAA TAC GGT TAT CCA CAG AAT CAG GCG ATA
 OCH tyr gly tyr pro gln asn gln gly ile
 8281/2761
 COT TGC TGG CGT TTT TCC ACA GGC TCC GCC
 arg cys trp arg phe ser ile gly ser ala
 8371/2791
 TAT AAA GAT ACC AGS CGT TTC CCC CTG GAA
 tyr lys asp thr arg arg phe pro leu glu
 8461/2821
 TCC CTT CCG GAA GCG TGG CCG TTT CTC ATA
 ser leu arg glu ala trp arg phe leu ile
 8551/2851
 ACG AAC CCC CCG TTC AGC CCG ACC GCT GCG
 thr asn pro pro phe ser pro thr ala ala
 8641/2881
 CAG CAG CCA CTG GTA ACA GGA TTA GCA GAG
 gln gln pro leu val thr gly leu ala glu
 8731/2911
 GAA GGA CAG TAT TTG GTA TCT GCG CTC TGC
 glu gly gln tyr leu val ser ala leu cys
 8821/2941
 CTG GTA GCG GTG GTT TTT TTG TTT GCA AGC
 leu val ala val val phe leu phe ala ser
 8911/2971
 CTG ACG CTC AGT GGA ACG AAA ACT CAC GTT
 leu thr leu ser gly thr lys thr his val
 9001/3001
 AAT GAA GTT TTA AAT CAA TCT AAA GTA TAT
 asn glu val leu asn gln ser lys val tyr
 9051/3031
 TCT GTC TAT TTC GTT CAT CCA TAG TTG CTT
 ser val tyr phe val his pro asn leu pro
 9151/3061
 CAA TGA TAC CCG GAG ACC CAC GCT CAC CCG
 gln opa tyr arg glu thr his ala his arg
 9271/3091
 CAA CTT TAT CCG CTT CCA TCC AGT CTA TTA
 gln leu tyr pro pro pro ser ser leu leu
 9361/3121
 CCA TTS CTA CAG GCA TCG TGG TGT CAC GCT
 pro leu leu gln ala ser trp cys his ala
 9451/3151
 CCC CCA TGT TGT GCA AAA AAG CCG TTA GCT
 pro pro cys cys ala lys lys arg leu ala
 9541/3181
 TCG CAG CAC TGC ATA ATT CTC TTA CTG TCA
 trp gln his cys ile ile leu leu leu ser
 9631/3211
 AGT GTA TGC GCG CAG CCA GTT GCT GCC
 ser val cys gly asp arg val ala leu ala
 9721/3241
 GAA AAC GTT CTT CCG GCG GAA AAC TCT CAA
 glu asn val leu arg gly glu asn ser gln
 9811/3271
 CAG CAT CTT TTA CTT TCA CCA GCG TTT CTG
 gln his leu leu leu ser pro ala phe leu
 9901/3301
 GTT GAA TAC TCA TAC TCT TCC TTT TTC AAT
 val glu tyr ser tyr ser ser phe phe asn
 9991/3331
 AGA AAA ATA AAC AAA TAG GCG TTC CCG GCA
 arg lys ile asn lys asn gly phe arg ala
 10081/3361
 ATA AAA ATA GCG GTA TCA CGA GCG CTT CTC
 ile lys ile gly val ser arg gly pro phe
 10171/3391
 TCA CAG CTT GTC TGT AAG CCG ATG CCG GGA
 ser gln leu val cys lys arg met pro gly
 7581/2581
 AAA CTC GAA AAT GTC TTA TAT TTC GAA GCA
 lys leu glu asn val leu tyr phe glu ala
 7771/2591
 CAC GAC GGC ACG TTG GAG TAC TTT AGT CCG
 his asp gly thr leu glu tyr phe ser pro
 7861/2621
 ACA AGT TGC TTA CCG GCG GTT COT AAT CAT
 thr ser cys OCH pro ala val arg asn his
 7951/2651
 CCG GAA GCA TAA AGT GTA AAG CCT GCG GTG
 pro glu ala OCH ser val lys pro gly val
 8041/2681
 GAA ACC TGT COT GCC AGC TCC ATT AAT GAA
 glu thr cys arg ala ser cys ile asn glu
 8131/2711
 CTG ACT CCG TGC GCT CCG TTG TTC GGC TGC
 leu thr arg cys ala arg ser phe gly cys
 8221/2741
 ACG CAG GAA ACA ACA TGT GAG CAA AAG GCG
 thr gln glu arg thr cys glu gln lys ala
 8311/2771
 GAA ACC TGT COT GCC AGC TCC ATT AAT GAA
 glu thr cys arg ala ser cys ile asn glu
 8401/2801
 GCT CCC TCG TGC GCT CTC CTG TTC CGA CCC
 ala pro ser cys ala leu phe arg pro pro
 8491/2831
 GCT CAC GGT GTA GGT ATC TCA GTT CCG TGT
 ala his ala val gly ile ser val arg cys
 8581/2861
 COT TAT CCG GTA ACT ATC GTC TTG AGT CCA
 pro tyr pro val thr ile val leu ser pro
 8671/2891
 CGA GGT ATG TAG CCG GTG CTA CAG AGT TCT
 arg gly met asn ala val leu gln ser ser
 8761/2921
 TGA ACG CAG TTA COT TTG GAA AAA GAG TTG
 opa ser gln leu pro ser glu lys glu leu
 8851/2951
 AGC AGA TTA CCG GCA GAA AAA AAG GAT CTC
 ser arg leu arg ala glu lys lys asp leu
 8941/2981
 AAG GGA GTT TCG TCA TGA GAT TAT CAA AAA
 lys gly phe trp ser opa asp tyr gln lys
 9031/3011
 ATG AGT AAA CTT GGT CTC ACA GTT ACC AAT
 met ser lys leu gly leu thr val thr val
 9121/3041
 GAC TCC CCG TCG TGT AGA TAA CTA CGA TAC
 asp ser pro ser cys arg OCH leu arg tyr
 9211/3071
 CTC CAG ATT TAT CAG CAA TAA ACC AGC CAG
 leu gln ile tyr gln gln OCH thr ser gln
 9301/3101
 ATT GTT GCC CCG AAG CTA GAG TAA GTA GTT
 ile val ala gly lys leu glu OCH val val
 9391/3131
 COT COT TTG GTA TGG CTT CAT TCA GCT CCG
 arg arg leu val trp leu his ser ala pro
 9481/3161
 OCT TCG GTC CTC CGA TCG TTG TCA GAA GTA
 pro ser val leu arg ser leu ser glu val
 9571/3191
 TGC CAT CCG TAA GAT GCT TTT CTG TGA CTG
 cys his pro OCH asp ala phe leu opa leu
 9661/3221
 CCG COT CAA TAC GCG ATA ATA CCG CCG CAC
 arg arg gln tyr gly ile ile pro arg his
 9751/3251
 GGA TCT TAC CCG TGT TGA GAT CCA GTT CGA
 gly ser tyr arg cys opa asp pro val arg
 9841/3281
 GGT GAG CAA AAA CAG GAA GCG AAA ATG CCG
 gly glu gln lys gln glu gly lys met pro
 9931/3311
 ATT ATT GAA GCA TTT ATC AGG GTT ATT GTC
 ile ile glu ala phe ile arg val ile val
 10021/3341
 CAT TTC CCG GAA AAG TGC CAC CTG ACC TCT
 his phe pro glu lys cys his leu thr ser
 10111/3371
 GTC TCG CCG GTT TCG GTG ATG AGG GTG AAA
 val ser arg val ser val met thr val lys
 10201/3401
 GCA GAC AAG CCC GTC AGG CCG COT CAG CCG
 ala asp lys pro val arg ala arg gln arg
 7711/2571
 CTT GAT CCG GTG TAT GTT TGC GAT TAC GGA
 leu asp arg val tyr val cys asp tyr gly
 7801/2601
 GAA AAA ATT CGA CAC ACA ACT ATG CAC GTT
 glu lys ile arg his thr thr met his val
 7891/2631
 GGT CAT AGC TGT TTC CTG TGT GAA ATT GTT
 gly his ser cys phe leu cys glu ile val
 7981/2661
 CCT AAT GAG TGA GCT AAC TCA CAT TAA TTG
 pro asn glu opa ala asn ser his OCH leu
 8071/2691
 TCG CCG AAC CCG CCG GGA GAG CCG GTT TCC
 ser ala asn ala arg gly glu ala val cys
 8161/2721
 GCG GAG CCG TAT CAG CTC ACT CAA AGG CCG
 gly glu arg tyr gln leu thr gln arg arg
 8251/2751
 ACC AAA ACG CCA GGA ACC GTA AAA AGG CCG
 ser lys arg pro gly thr val lys arg pro
 8341/2781
 CAA GTC AGA GGT CCG GAA ACC CGA CAG GAC
 gln val arg gly gly glu thr arg gln asp
 8431/2811
 TCC CCG TTA CCG GAT ACC TGT CCG CTT TTC
 thr cys leu pro asp thr cys pro pro phe
 8521/2841
 AGG TCG TTC GCT CCA AGC TCG GCT GTG TCC
 arg ser phe ala pro ser trp ala val cys
 8611/2871
 ACC CCG TAA GAC ACG ACT TAT CCG CAC TCG
 thr arg OCH asp thr thr tyr arg his trp
 8701/2901
 TGA AGT GGT GCG CTA ACT ACG GCT ACA CTA
 opa ser gly gly leu thr thr ala thr leu
 8791/2931
 GTA OCT CTT GAT CCG CCA AAC AAA CCA CCG
 val ala leu asp pro ala asn lys pro pro
 8881/2961
 AAG AAG ATC CTT TGA TCT TTT CTA CCG GGT
 lys lys ile leu opa ser phe leu arg gly
 8971/2991
 GCA TCT TCA COT AGA TCC TTT TAA ATT AAA
 gly ser ser pro arg ser phe OCH ile lys
 9061/3021
 GCT TAA TCA CTG AGG CAC CTA TCT CAG CGA
 ala OCH ser val arg his leu ser gln arg
 9151/3051
 CCG AGG GCT TAC CAT CTG GCC CCA GTG CTG
 gly arg ala tyr his leu ala pro val leu
 9241/3081
 CCG GAA GCG CCG AGC GCA GAA GTG GTC CTG
 pro glu gly pro ser ala glu val val leu
 9331/3111
 CCG CAG TTA ATA GTT TCC GCA ACG TTG TTG
 arg gln leu ile val cys ala thr leu leu
 9421/3141
 GTT CCC AAC GAT CAA GCG GAG TTA CAT GAT
 val pro asn asp gln gly glu leu his asp
 9511/3171
 AGT TCG CCG CAG TGT TAT CAC TCA TCG TTA
 ser trp pro gln cys tyr his ser trp leu
 9601/3201
 GTG AGT ACT CAA CCA AGT CAT TCT GAG AAT
 val ser thr gln pro ser his ser glu asn
 9691/3231
 ATA GCA GAA CTT TAA AAG TGC TCA TCA TTG
 ile ala glu leu OCH lys cys ser ser leu
 9781/3261
 TGT AAC CCA CTC GTG CAC CCA ACT GAT CTT
 cys asn pro leu val his pro thr asp leu
 9871/3291
 CAA AAA AGG GAA TAA GCG CGA CAC GGA AAT
 gln lys arg glu OCH gly arg his gly asn
 9961/3321
 TCA TGA GCG GAT ACA TAT TTG AAT GTA TTT
 ser opa ala asp thr tyr leu asn val phe
 10051/3351
 AAG AAA CCA TTA TTA TCA TGA CAT TAA CCT
 lys lys pro leu leu ser opa his OCH pro
 10141/3381
 ACC TCT GAC ACA TGC AGC TCC CCG AGA CCG
 thr ser asp thr cys ser ser arg arg arg
 10231/3411
 GTG TTG CCG GGT GTC GCG GCT GCG TTA ACT
 val leu ala gly val gly ala gly leu thr

pACgp67-ScFv350 -> 1-phase Translation

Fig 2E

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10251/3421	10291/3431	10321/3441
ATG CCG CAT CAG AGC AGA TTG TAC TGA GAG TGC ACC ATA TGC GGT GTG AAA TAC CCG ACA GAT GCG TAA GGA GAA AAT ACC GCA TCA GGC		
met arg his gln ser arg leu tyr opa glu cys thr ile cys gly val lys tyr arg thr asp ala och gly glu asn thr ala ser gly		
10351/3451	10381/3461	10411/3471
GCC ATT CCG CAT TCA GGC TGC GCA ACT GTT GGG AAG GGC GAT CCG TGC GGG CCG CTT CCG TAT TAC GCC AGC TGG CGA AAG GGG GAT GTG		
ala ile arg his ser gly cys ala thr val gly lys gly asp arg cys gly pro leu arg tyr tyr ala ser trp arg lys gly asp val		
10441/3481	10471/3491	10501/3501
CTG CAA GGC GAT TAA GTT GGG TAA CCG CAG GGT TTT CCC AGT CAC GAC GTT GTA AAA CGA CCG CCA GTG CC		
leu gln gly asp och val gly och arg gln gly phe pro ser his asp val val lys arg arg pro val		

Figure 3 : clonetherap.99B3

VH sequence :

GAGGTGAAGCTTCTCCAGTCTGGAGGTGGCCTGGTGCAGCCTGGAGGATCCCTGA
AACTCTCCTGTGCAGCCTCAGGAATCGATTTTAGTAGATACTGGATGAGTTGGGT
TCGGCGGGCTCCAGGGAAAGGACTAGAATGGATTGGAGAAATTAATCCAGATAG
CAGTACAATAAACTATGCACCATCTCTAAAGGATAAAATTCATCATCTCCAGAGAC
AACGCCAAAAATACGCTGTACCTGCAAATGAGCAAAGTGAGATCTGAGGACACA
GCCCTTTATTACTGTGCAAGAGGACTGGGACAGAACTTTGACTACTGGGGCCAAG
GCACCACTCTCACAGTCTCCTCA

VL sequence :

GATATTGTGATGACGCAGGCTGCATTCTCCAATCCAGTCACTCTTGGAACATCAG
CTTCCATCTCCTGCAGGTCTAGTAAGAGTCTCCTACATAGTAATGGCATCACTTAT
TTGTATTGGTATCTGCAGAAGCCAGGCCAGTCTCCTCAGCTCCTGATTTATCAGAT
GTCCAACCTTGCCCTCAGGAGTCCCAGACAGGTTCAGTAGCAGTGGGTCAGGAACT
GATTTACACTGAGAATCAGCAGAGTGGAGGCTGAGGATGTGGGTGTTTATTACT
GTGCTCAAAATCTAGAACTTCCGTGGACGTTCCGGTGGAGGCACCAAGCTGGAAAT
CAAA

Figure 4 : clonetherap.88E10

VH sequence :

GAGGTGAAGCTGGTGGAGTCTGGAGGAGGCTTGGTACAGCCTGGGGGTCTCTG
AGTCTCTCCTGTGCAGCTTCTGGATTACCTTCACTGATTACTCCATGAACTGGGT
CCGCCAGCCTCCAGGGAAGACACTTGAGTGGTTGGCTTTTATTAGAAACAAAGCT
AATGGTTACACAGCAGAGTACAGTGCATCTGTGAAGGGTCGGTTCTCCATCTCCA
GAGATAATTCCCAAAGCATCCTCTATCTTCAAATGAATGCCCTGAGAGCTGAGGA
CAGTGCCACTTATTACTGTGCAAGGGGATGGTATGCTATGGACTACTGGGGTCAA
GGAACCTCAGTCACCGTCTCCTCA

VL sequence :

Figure 5 : clonetherap.152C3

VH sequence :

GAGGTTCTGCTGCAGCAGTCTGTGGCAGAGCTTGTGAGGCCAGGGGCCTCAGTCA
AGTTGTCCTGCATAGTTTCTGACTTCAACATTAAACACACCTATATGCACTGGGTG
AAACAGAGGCCTGATCAGGGCCTGGAGTGGATTGGAAGGATTGATCCTGCGAAT
GGTAAAACTATATATGCCCCGACGTTCCAGGGCAAGGCCACTATAACTGCGGACA
CATCCTCCAACACAGCCTACCTGCATTTCAGCAGCCTGACATCTGAGGACGCTGC
CATCTATTACTGTGCTAGAGCTGGGGCTGGCTACTTTGACTACTGGGGCCAAGGC
ACCACTCTCACAGTCTCCTCA

VL sequence :

GACATCTTGCTGACTCAGTCTCCAGCCATCCTGTCTGTGAGTCCAGGAGAAAGAG
TCAGTTTCTCCTGCAGGGCCAGTCAGAACATTGGCACAAGTATTTACTGGTATCA
GCAAAGAACAAATGGTTCTCCAAGGCTTCTCATAAAGTATGTTTCTGAGTCTATC
TCTGGGATCCCTTCCAGGTTTAGTGGCAGTGGATCAGGGACAGAGTTTACTCTTA
GCATCAACAGTGTGGAGTCTGAAGATATTGCAGATTATTACTGTCAACAAAGTCA
TAGTTGGCCGCTCACGTTCTGTGCTGGGACCAAGCTGGAGCTGAAA

Fig 6

ToxO
Interaction ANT-Vpr52-96

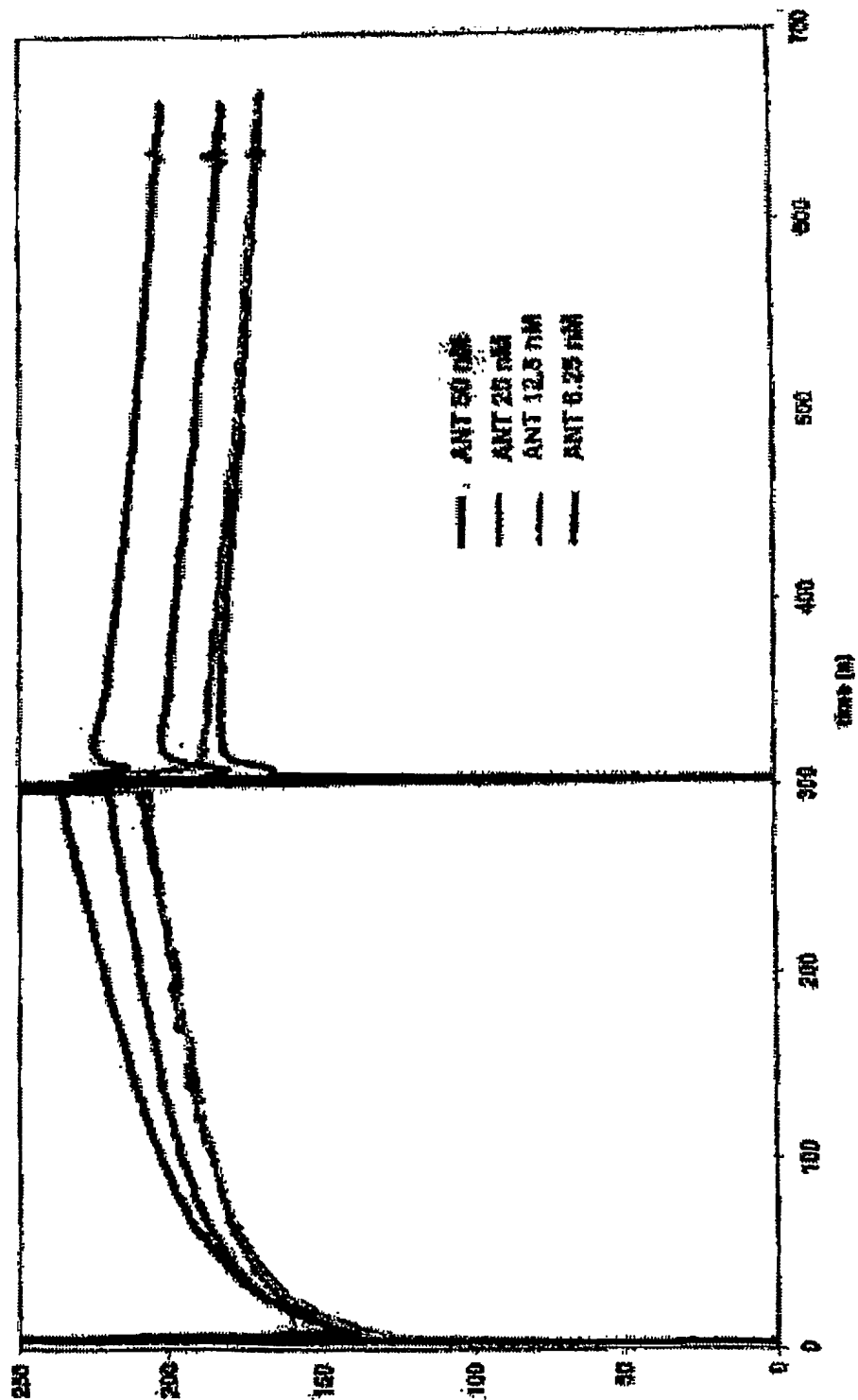


Fig. 7

Interaction ANT-Tox1

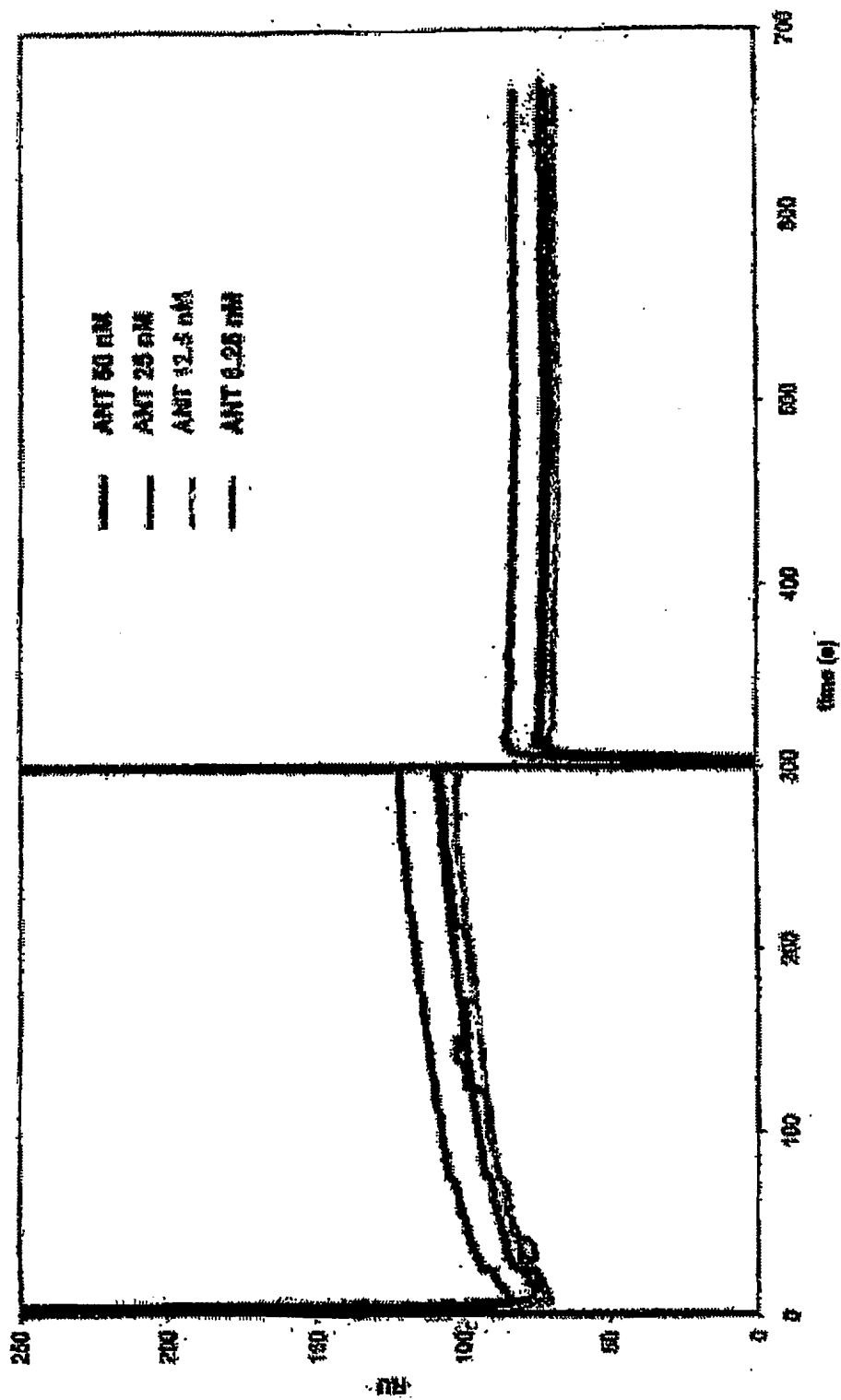


Fig. 8

Tox0
Interaction VDAC-Wpr52-96

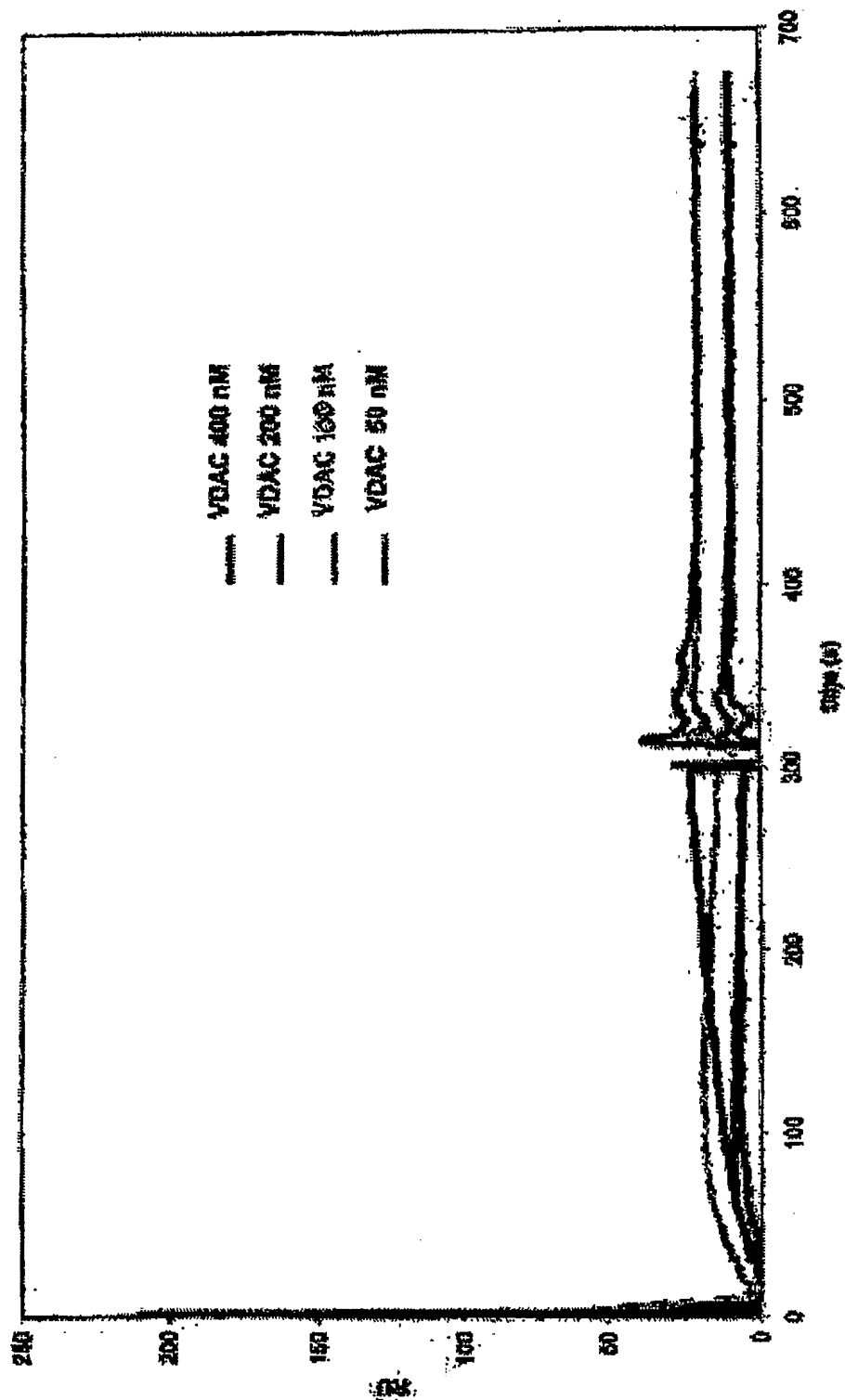


Fig 9

Interaction VDAC-Tox1

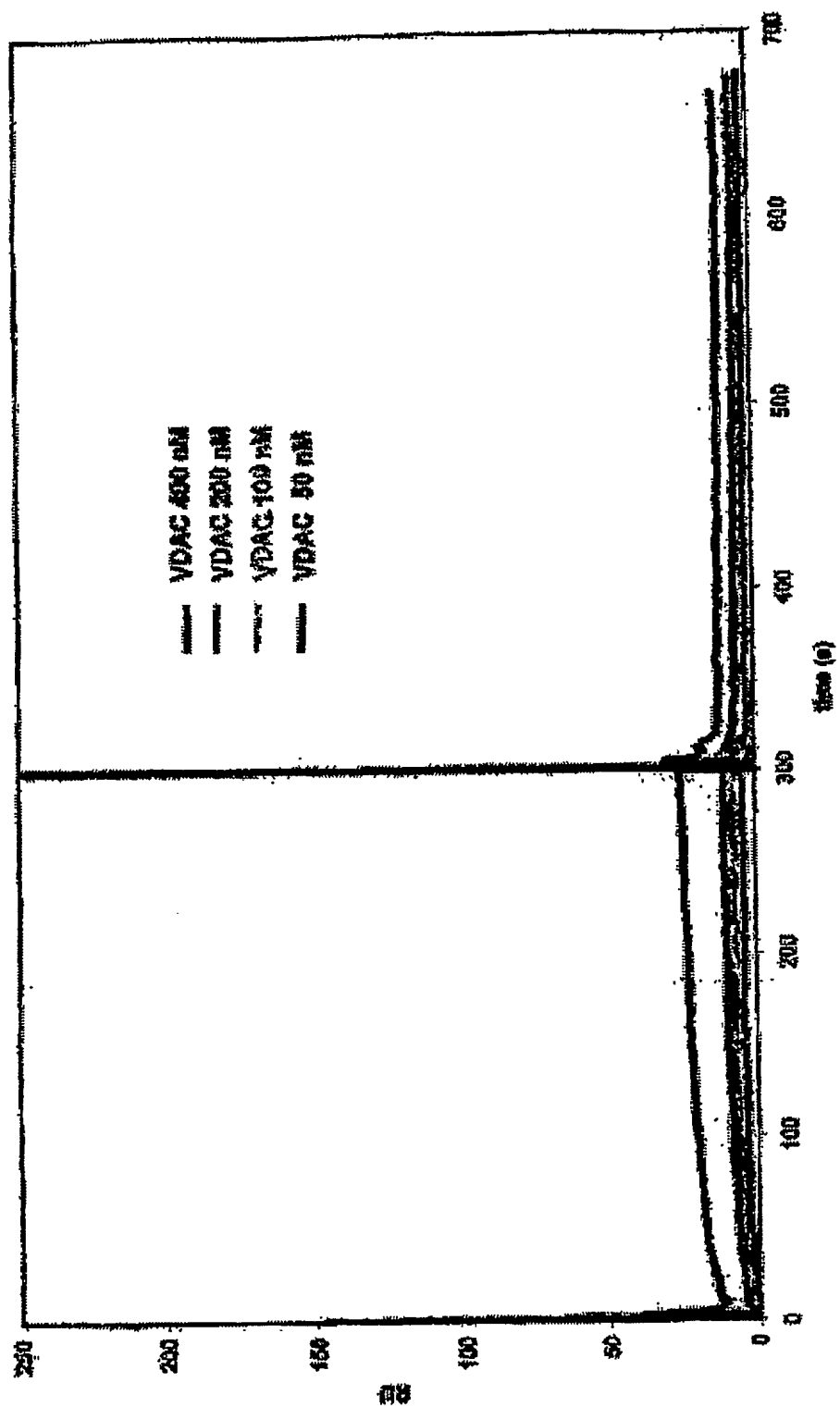


Fig. 10

Interaction ANT 80 nM - peptides

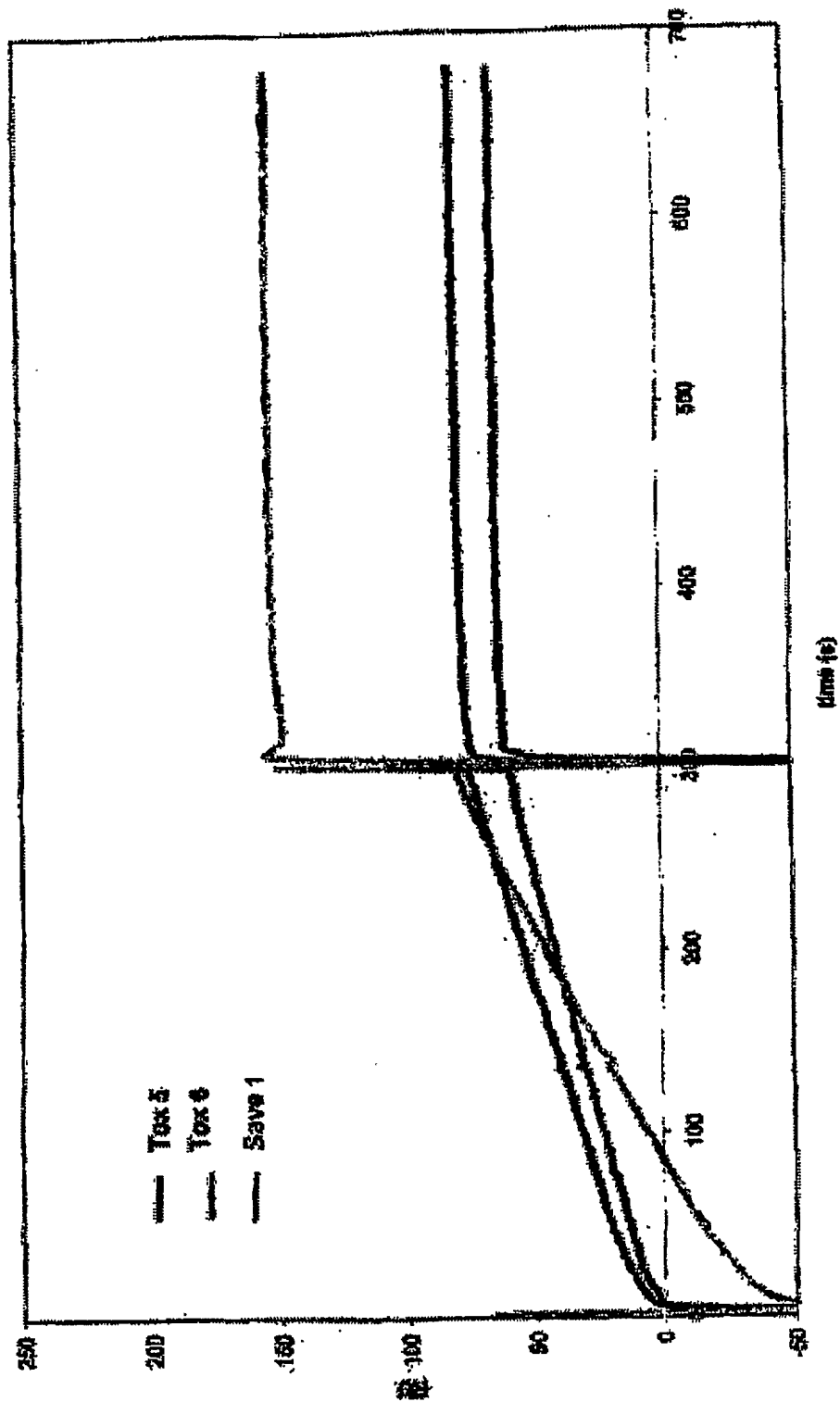
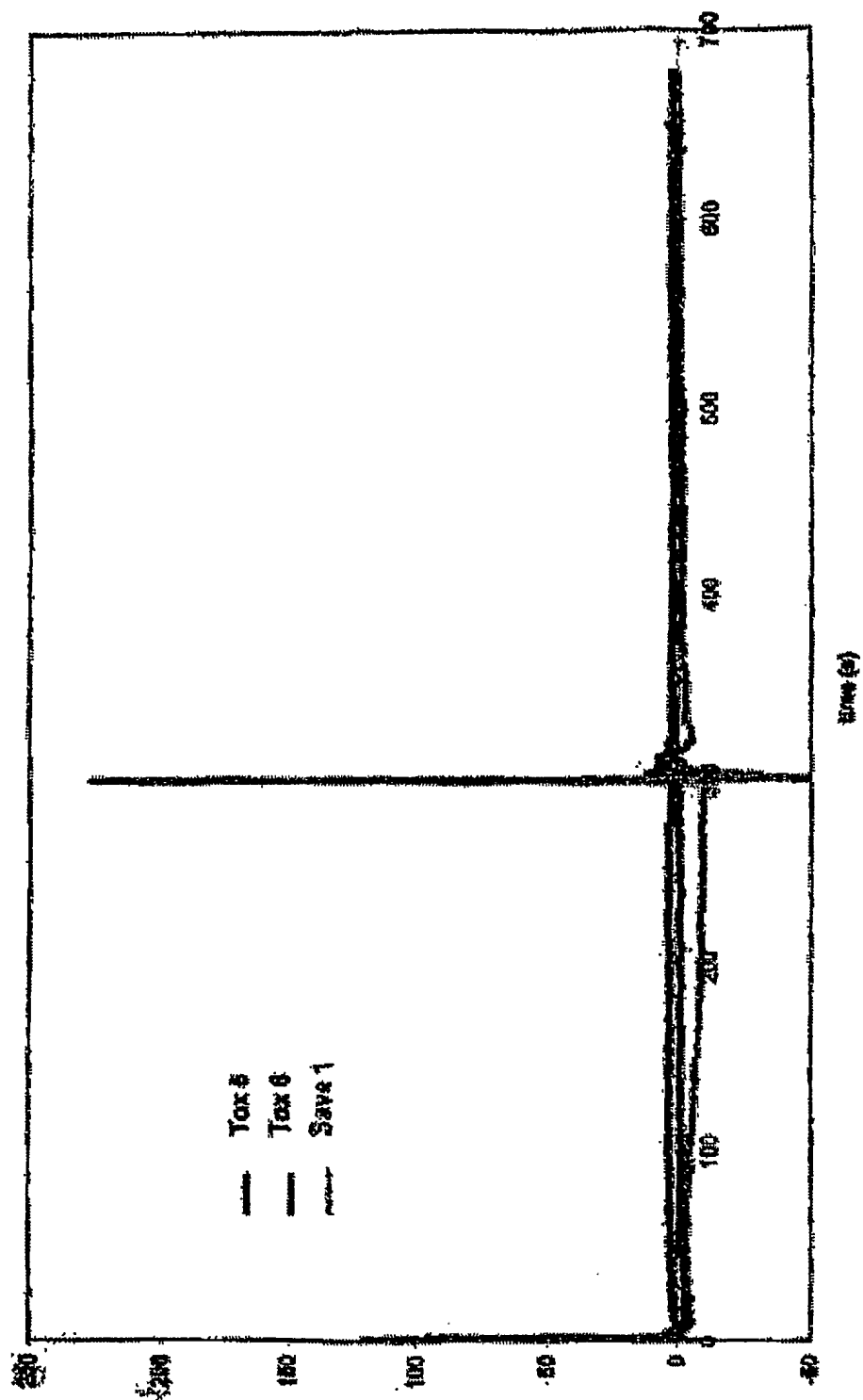


Fig. 11

Information VPAC 50 nM - peptides



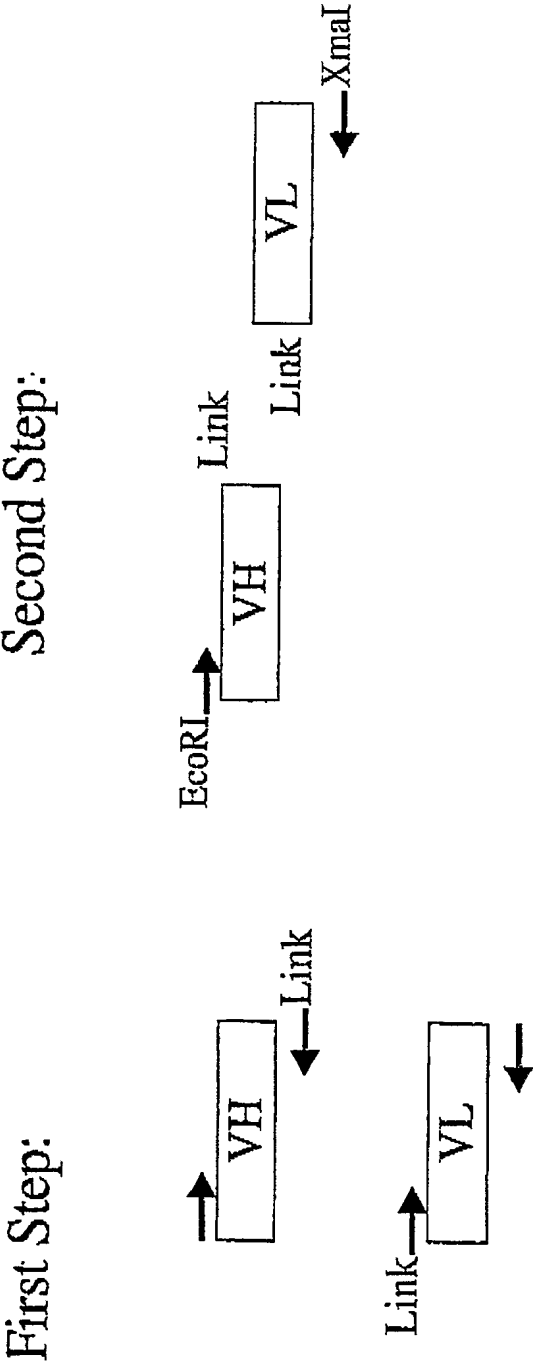


Figure 12: Obtention of the VH/VL chimeric DNA

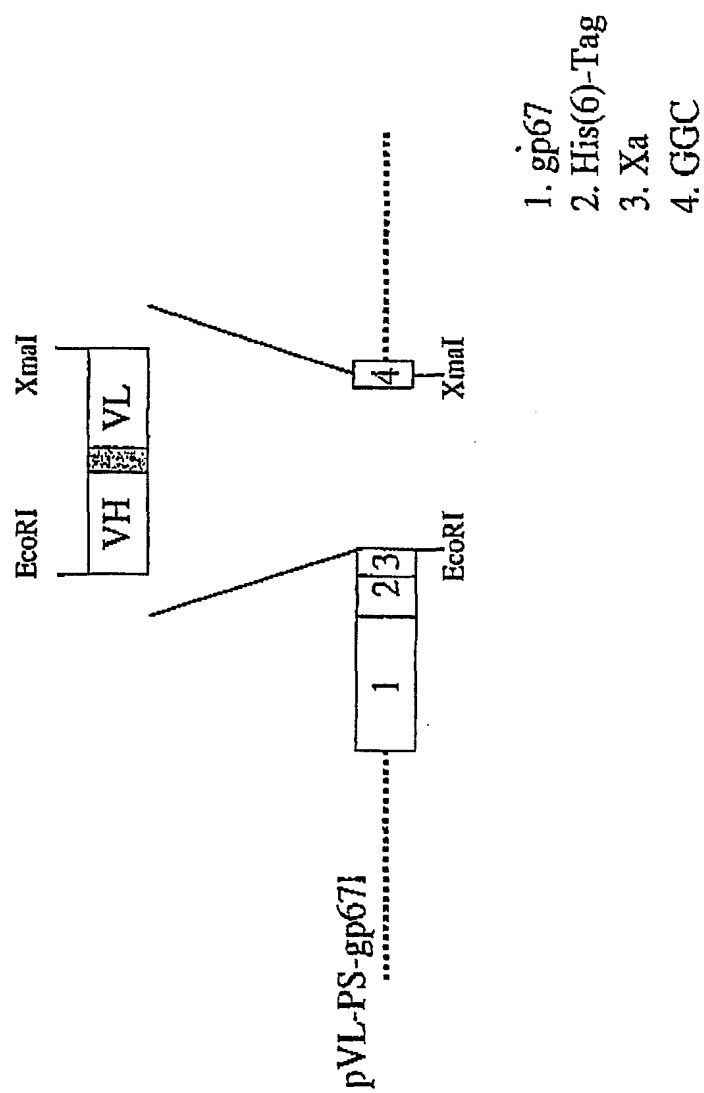


Figure 13: Map of the ScFv transfer vector